

RESEARCH

Open Access

Lipid peroxidation and antioxidant enzymes activity in *Plasmodium vivax* malaria patients evolving with cholestatic jaundice

Camila Fabbri¹, Rita de Cássia Mascarenhas-Netto², Pritesh Lalwani^{1,5}, Gisely C Melo^{3,4}, Belisa ML Magalhães^{3,4}, Márcia AA Alexandre^{3,4}, Marcus VG Lacerda^{3,4*} and Emerson S Lima¹

Abstract

Background: *Plasmodium vivax* infection has been considered a benign and self-limiting disease, however, recent studies highlight the association between vivax malaria and life-threatening manifestations. Increase in reactive oxygen species has already been described in vivax malaria, as a result of the increased metabolic rate triggered by the multiplying parasite, and large quantities of toxic redox-active byproducts generated. The present study aimed to study the oxidative stress responses in patients infected with *P. vivax*, who developed jaundice (hyperbilirubinaemia) in the course of the disease, a common clinical complication related to this species.

Methods: An evaluation of the lipid peroxidation and antioxidant enzymes profile was performed in 28 healthy individuals and compared with *P. vivax* infected patients with jaundice, i.e., bilirubin < 51.3 µmol/L (8 patients) or without jaundice (34 patients), on day 1 (D1) and day 14 (D14) after anti-malarial therapy.

Results: Hyperbilirubinaemia was more frequent among women and patients experiencing their first malarial infection, and lower haemoglobin and higher lactate dehydrogenase levels were observed in this group. Malondialdehyde levels and activity of celuroplasmín and glutathione reductase were increased in the plasma from patients with *P. vivax* with jaundice compared to the control group on D1. However, the activity of thioredoxin reductase was decreased. The enzymes glutathione reductase, thioredoxin reductase, thiols and malondialdehyde also differed between jaundiced versus non-jaundiced patients. On D14 jaundice and parasitaemia had resolved and oxidative stress biomarkers were very similar to the control group.

Conclusion: Cholestatic hyperbilirubinaemia in vivax malaria cannot be totally disassociated from malaria-related haemolysis. However, significant increase of lipid peroxidation markers and changes in antioxidant enzymes in patients with *P. vivax*-related jaundice was observed. These results suggest oxidative processes contributing to malaria pathogenesis, what may be useful information for future anti-oxidant therapeutical interventions in these patients.

Keywords: Malaria, *Plasmodium vivax*, Antioxidant enzymes, Oxidative stress, Jaundice, Hyperbilirubinaemia

Background

Malaria affects millions of people every year around the world [1]. *Plasmodium falciparum* is the most lethal species responsible for the major burden of malaria disease in Africa. However, *Plasmodium vivax* is the most abundantly distributed species worldwide. Recent

reports suggest increasing clinical complications in *P. vivax* infected individuals in many endemic regions [2,3]. Brazil reports ~50% of the malarial cases in the Americas and approximately 99.5% of these cases occur in the Amazon Region [4]. Some data suggest an increased rate of hospitalization due to *P. vivax* infection in the Brazilian Amazon region over the past years [5]. Part of this increased hospitalization is related to side effects of anti-malarial drugs, such as primaquine (used as anti-hypnozoiticidal to avoid relapses), leading to haemolysis in patients with glucose-6-phosphate-dehydrogenase

* Correspondence: marcuslacerda.br@gmail.com

³Fundação de Medicina Tropical Dr. Heitor Vieira Dourado, Manaus, AM 69040-000, Brazil

⁴Universidade do Estado do Amazonas, Manaus, AM 69040-000, Brazil

Full list of author information is available at the end of the article

(G6PD) deficiency [6]. Nevertheless, a considerable proportion of patients develop clinical complications very similar to those observed in *P. falciparum* severe disease, such as jaundice, anaemia, acute renal failure, shock and coma, being the first the most common [7-10], and in most of these reports, *P. vivax* mono-infection is confirmed by PCR.

Severity criteria for *P. falciparum* are relatively well-established in the literature [11], and recently the same criteria are being used by several authors for severe *P. vivax* disease. Particularly, jaundice (impregnation of soft tissues by elevated bilirubins) in malaria may be explained by severe haemolysis (indirect bilirubin predominance) or liver cholestasis (direct bilirubin predominance). World Health Organization (WHO) most recent recommendations state that jaundice by itself (serum total bilirubins > 51.3 $\mu\text{mol/L}$) should not be used as a single marker of severity *sensu strictu*, because it is not associated with higher fatality rates, unless there are other simultaneous organs dysfunctions [12,13]. Recent data from the Brazilian Amazon have also shown that hyperbilirubinaemia is not independently associated to intensive care unit hospitalization of children with vivax malaria [14]. The prognosis is favourable, and jaundice vanishes in parallel with peripheral parasitaemia clearance. However, malarial infection causing hyperbilirubinaemia with clinical jaundice leads to persistent vomiting, and is a major cause of prolonged hospitalization in many sites where *P. vivax* is endemic, contributing to increase the social and economic burden of this disease [13].

Despite the frequent occurrence of hyperbilirubinaemia, very little progress has been made in understanding the pathogenesis of cholestasis jaundice in patients with malaria, particularly in vivax disease. Increase in reactive oxygen species (ROS) has already been described in vivax malaria. As a result of the increased metabolic rate of the rapidly growing and multiplying parasite, large quantities of toxic redox-active byproducts are generated. Furthermore, a reduction in antioxidant enzymes such as glutathione peroxidase, catalase and superoxide dismutase has been observed in plasma of malaria-infected individuals [15-17]. These changes in oxidants and anti-oxidants have been associated with severe malaria in children [18]. Oxidative stress (OS) in malaria can be caused by two main mechanisms. Firstly, by the parasite, which reproduces in the erythrocytes, changing the structure and affecting parameters such as stiffness, viscosity and volume. Central to the generation of OS is the degradation of host haemoglobin by the parasite. Secondly, the OS mechanisms involve the host immune response, which initiates a cascade of defense mechanisms culminating with the release of free radicals by activated macrophages, to tackle the parasite [19,20]. Furthermore, reactive hydroxyl radicals ($\cdot\text{OH}$) generated via mitochondrial OS, have been shown to play

an important role in the liver apoptosis in a murine model of malarial infection [21,22].

Based on previous studies demonstrating the role of OS upon other clinical complications of *P. vivax* infection, it was therefore hypothesized that the transitory predominantly cholestatic jaundice seen in vivax malaria could also be associated to OS.

Methods

Study design

Patients with any clinical complications attributed to malaria are systematically hospitalized in the Clinical Investigation Ward of the *Fundação de Medicina Tropical Dr. Heitor Vieira Dourado* (FMT-HVD), a reference tertiary care center for infectious diseases located in Manaus (Western Brazilian Amazon). In this ward, the staff completed a standard questionnaire regarding epidemiological and clinical characteristics of the patients. Blood samples were collected before the beginning of the routine anti-malarial treatment with chloroquine (25 mg/kg over 3 days) and primaquine (0.5 mg/kg/day for 7 days), according to the National Anti-malarial Guidelines. Healthy volunteers without past history of malaria served as controls.

Patients included in this study had no diabetes or arterial hypertension history (as confirmed by fast glucose and arterial tension repeated measures throughout the hospitalization period), and were systematically phenotyped for G6PD deficiency, according to the technique described elsewhere [23]. G6PD deficient patients were not included in the analysis. In all these patients, *P. vivax* mono-infection was confirmed by PCR [24], ruling out mixed infections with *P. falciparum*. Other common infectious diseases leading to cholestasis were also ruled out through specific antibody detection (leptospirosis, hepatitis A, hepatitis B, hepatitis C and HIV), blood culture (bacterial infection), and RT-PCR (dengue virus 1,2,3 and 4). Abdominal ultrasound was also performed in all patients to exclude lithiasic cholecystitis or any other biliary tract abnormality. On day 14 (D14) after the beginning of therapy (D1), patients were informed to return to the Outpatient Clinics for clinical and laboratorial re-evaluation. Thick blood smear with parasitaemia count in 100 leukocytes, automatized full blood count and serum biochemical analysis (aspartate aminotransferase - AST, alanine aminotransferase - ALT, alkaline phosphatase - AP, gamma-glutamyltransferase - gamma-GT, bilirubins, lactic dehydrogenase - LDH) were systematically performed on D1 and D14.

Blood samples

About 15 mL of venous blood were collected on BD Vacutainer® tubes with and without $\text{K}_2\text{-EDTA}$. Aliquots of plasma were stored at -70°C before analysis.

Oxidative stress biomarkers

Malondialdehyde (MDA) (a marker of free radical activity and lipid peroxidation) was measured using a spectrophotometer 70 UV/VIS Spectrometer PG Instruments Ltda (Beijing, China) by reaction with thiobarbituric acid (TBA) in plasma [25]. Glutathione reductase (GR; E.C. 1.6.4.2) was measured in plasma using Randox® kits on a microplate reader DTX 800 Multimode Detector, Beckman Coulter (Fullerton, CA, USA). The activity of the enzyme thioredoxin reductase (TrxR; E.C. 1.8.1.9) [26] and ceruloplasmin (CP; E.C. 1.16.3.1) [27] was performed in plasma by microplate readers. Thiol compounds were measured in plasma using the modified method [28,29] where 300 µL of 0.25 mM Tris + 20 mM EDTA pH 8.2, 3,8 µL of 5,5-dithiobis acid-2-nitrobenzoic (DTNB) 0.1 M and 7,5 µL of standard (0.5 mM glutathione) sample or water (blank) were incubated at room temperature for 15 minutes and measured in a microplate reader at a wavelength of 412 nm. All chemicals and reagents used in the study were purchased from Sigma-Aldrich® (St. Louis, MO, USA) and Randox® kits (County Antrim, UK).

Ethical approval

The study was approved by the FMT-HVD Ethics Review Board (CAAE-0075.0.115.114-11), and all the patients signed a written consent after being informed about the objectives of the study.

Statistical analysis

Normal distribution was assessed through Shapiro-Wilk test. Parametric data were analysed by ANOVA-one way to estimate mean differences. When significant, *post-hoc* Tukey test was performed. Kruskal-Wallis test was used for non-parametric analysis. Student and Mann-Whitney tests were used when only two groups were compared. Frequency differences were detected using chi-square. Correlations between variables were performed using the Spearman test. All tests were performed in BioStat 5.0® (Universidade Federal do Pará, Belém, Brazil) and OriginPro 8.0® (Microcal, Northampton, Massachusetts, USA), and significance was considered when $p < 0.05$.

Results

During the year of 2011, 25 hospitalized patients were enrolled with confirmed microscopic diagnosis of *P. vivax* mono-infection, presenting with serum total bilirubin higher than 51.3 µmol/L (3.0 mg/dL) (direct bilirubin higher than indirect bilirubin, characterizing cholestasis). During the same period, 42 patients with non-complicated vivax malaria in the same age group, with bilirubin within the normal range, were also randomly enrolled. After ruling out other comorbidities

(lithiasic cholecystitis in 4, G6PD deficiency in 2, dengue fever in 5, chronic hepatitis B in 2, chronic hepatitis C in 1, HIV in 1 and Pf/Pv mixed infection by PCR in 2), a total of eight patients with vivax-related jaundice, 34 vivax patients without jaundice and 28 healthy volunteers were included in the final analysis. No complication other than hyperbilirubinaemia was observed after detailed clinical and laboratorial screening. On D14 a clinical and laboratorial screening was performed on seven out of eight with jaundice, and 18 out of 34 patients without jaundice. None of them presented with persistent parasitaemia, clinical jaundice or laboratory hyperbilirubinaemia on D14. None of the controls on D1 referred any clinical complication in between D1 and D14. Epidemiological, haematological and biochemical data are detailed in Table 1. Jaundice was more frequent among women and those experiencing malarial infection for the first time. Haemoglobin was lower in those with jaundice, and the levels of LDH, AST and ALT were higher in this group.

Oxidative stress biomarkers

A significant increase in MDA levels on D1 in *P. vivax* malaria (with and without jaundice) group was observed compared to the control group. In addition, a significant increase of MDA was observed on D1 in the jaundiced group compared to the non-jaundiced group (Figure 1).

Figure 2 shows altered antioxidant enzyme profile in malaria patients. CP and GR are significantly increased in malaria-infected individuals (with or without jaundice) on D1 (Figures 2A and 2B) and TrxR is lower in infected patients (Figure 2C), compared to healthy volunteers. Differences in GR, TrxR and thiols between jaundiced and non-jaundiced patients are also seen (Figures 2B, 2C and 2D). On D14, markers of oxidative stress were not different from the healthy volunteers group, suggesting a convalescent state after full clinical recovery (Figure 2).

Despite of the lower level of haemoglobin in the jaundiced group, no single plasmatic oxidative stress marker was correlated with haemoglobin levels (data not shown).

Discussion

A case series reported in 2010 showed that over 50% of patients admitted to a reference hospital in the city of Manaus had jaundice as complication for *P. vivax* malaria [8]. In the present study, evidence that *P. vivax* infection increases lipid peroxidation and alters profile of antioxidant enzymes is provided. That may eventually lead to hyperbilirubinaemia, which is not independently associated to severity [14], but was already reported in deceased patients with the clinical diagnosis of vivax malaria and other comorbidities [30]. In the eight jaundiced patients presented in this manuscript, however,

Table 1 Epidemiological, haematological and biochemical parameters of healthy volunteers and vivax malaria patients with and without jaundice (total bilirubin > 51.3 μmol/L) on Day 1

	Healthy volunteers (n = 28)	<i>P. vivax</i> malaria without jaundice (n = 34)	<i>P. vivax</i> malaria with jaundice (n = 8)	P-value*
Age	25.4 ± 4.6	37.6 ± 15.2	33.5 ± 14.0	0.4867
Male sex (%)	32	82	38	0.0316
First malarial infection (%)	-	24	63	0.0316
Time of disease (in days)	-	6.5 ± 2.7	9.7 ± 7.9	0.3960
Peripheral parasitaemia/μL	-	2,428 ± 4,276	5,353 ± 10,598	0.3578
Leukocytes (×10 ³ /μL)	6.69 ± 1.65	5.33 ± 1.59	5.41 ± 1.18	0.8903
Haemoglobin (g/dL)	13.0 ± 1.4	12.8 ± 1.8	10.8 ± 1.9	0.0051
Platelets (×10 ³ /μL)	228.5 ± 62.0	93.7 ± 52.0	85.1 ± 54.2	0.6790
TB (μmol/L)	18.1 ± 8.5	26.3 ± 10.1	109.3 ± 81.9	< 0.0001
IB (μmol/L)	10.1 ± 6.2	15.7 ± 7.2	28.6 ± 15.2	0.0547
DB (μmol/L)	8.0 ± 6.7	10.6 ± 6.3	80.7 ± 79.3	0.0003
AST (U/L)	21.5 ± 8.9	34.5 ± 17.4	74.7 ± 67.1	0.0488
ALT (U/L)	16.4 ± 8.7	34.5 ± 27.0	93.0 ± 91.8	0.0420
AP (U/L)	57.6 ± 21.4	72.4 ± 28.0	103.9 ± 68.2	0.2408
Gamma-GT (U/L)	11.5 ± 5.5	45.7 ± 37.3	106.4 ± 92.1	0.2408
LDH (U/L)	255.6 ± 74.3	425.2 ± 188.5	609.7 ± 234.8	0.0294

TB total bilirubin, IB indirect bilirubin, DB direct bilirubin, AST aspartate aminotransferase, ALT alanine aminotransferase, AP alkaline phosphatase, gamma-GT gamma-glutamyltransferase, LDH lactate dehydrogenase.

*Comparison between vivax malaria patients with and without jaundice. Significant values in bold. Continuous variables are presented as mean ± SD.

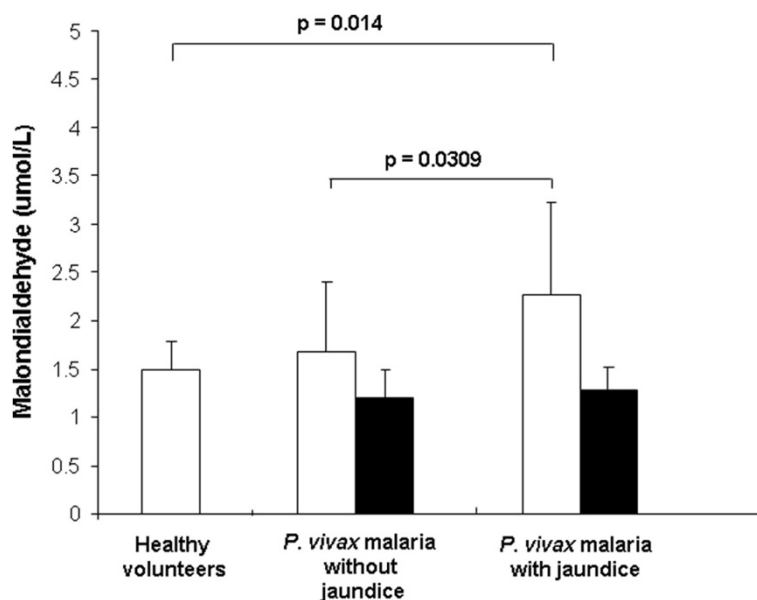


Figure 1 Lipid peroxidation levels expressed as malondialdehyde (MDA) in plasma on D1 (white bars) and D14 (black bars). Groups: Healthy volunteers (n = 28), *P. vivax* malaria without jaundice [n = 34 (D1); n = 18 (D14)] and *P. vivax* malaria with jaundice [n = 8 (D1); n = 7 (D14)]. Significant differences between the groups are indicated in each graph.

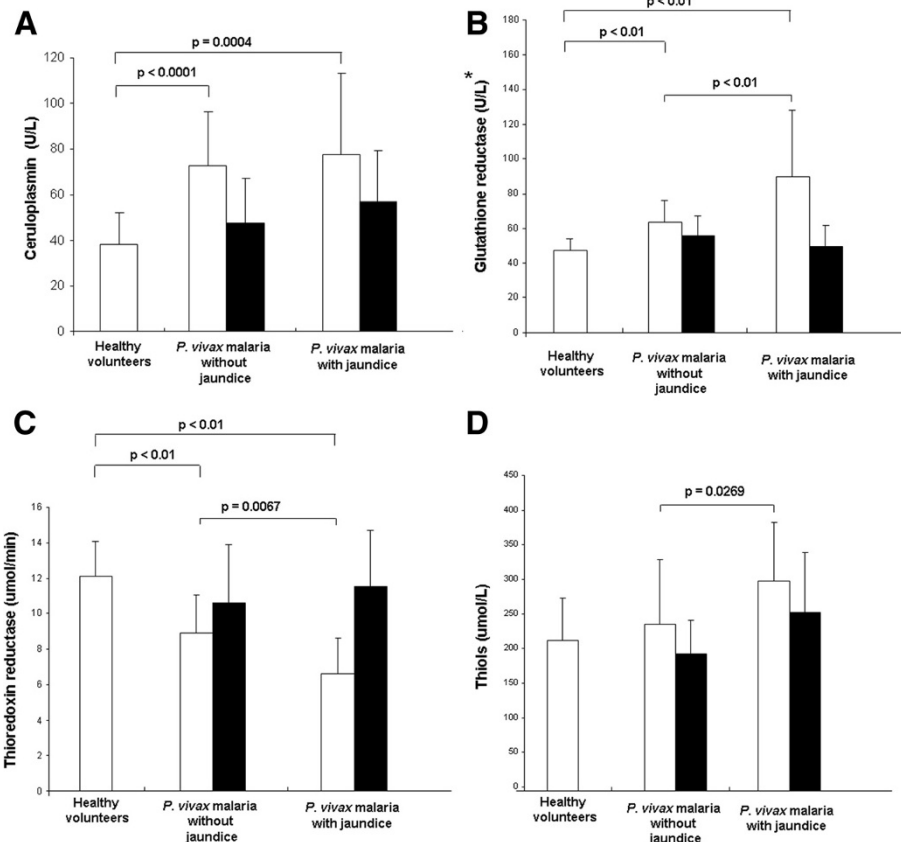


Figure 2 Antioxidant enzyme activities in plasma on D1 (white bars) and D14 (black bars). (A) Ceruloplasmin; (C) Thioredoxin reductase; (D) Thiols; healthy volunteers (n=28), *P. vivax* malaria without jaundice [n=34 (D1); n=18 (D14)] and *P. vivax* malaria with jaundice [n=8 (D1); n=7 (D14)]. Significant differences between the groups are indicated in each graph. *(B) Glutathione reductase: healthy volunteers (n=18), *P. vivax* malaria without jaundice [n=19 (D1); n=10 (D14)] and *P. vivax* malaria with jaundice [n=5 (D1); n=5 (D14)].

further comorbidities were ruled out by detailed laboratory screening.

Despite of the very small sample size, the finding of this complication more often in females contradicts previous findings showing that male children present with more life-threatening complications [11]. No obvious explanation for this phenomenon is known, except the hypothesis that more bile duct diseases are found in women, what may contribute to the cholestatic event. Higher occurrence in patients infected for the first time suggests an adaptation mechanism that needs to be explored with bigger sample sizes and prospective study designs.

The significant increased levels of MDA in plasma of malarial patients with and without jaundice may indicate the damage triggered by free radicals produced against cell membranes, such as erythrocytes and hepatocytes [31]. Recent studies have shown increased levels of lipid peroxidation markers such as MDA, which have been implicated in a number of diseases, including malaria [16,18,31]. *Plasmodium* sp. does not have a triglyceride synthesis pathway; therefore, it obtains all the lipids from

the host serum. The parasite also destabilize the cell membrane, in particular the erythrocyte membrane in order to obtain lipid [17,32].

The antioxidant enzyme TrxR activity was significantly decreased when patients with malaria were compared to the control group. TrxR catalyzes the reduction of the active site disulfide using NADPH. The high levels of *NO are associated with the host-parasite interaction. TrxR was shown to be inactivated with the production of nitric oxide (*NO), a reactive species of oxygen found in high levels during malaria infection [33]. This may partially explain the decreased levels of this enzyme in vivax patients.

On the other hand, the antioxidant enzymes GR and CP activities were significantly increased in *P. vivax* infected patients (with or without jaundice) compared with the control group. Other studies have also demonstrated increased levels of the enzyme GR in malaria caused by *Plasmodium berghei* and *P. falciparum* [19]. GR is involved in maintaining an intracellular reducing environment, which is crucial to the cell in the defense

against oxidative stress. Thus, increased levels of GR may be playing a role in counteracting with increased oxidant species and maintaining homeostasis [34]. Recent reports are in line with these results, confirming increased CP activity in malaria [35,36]. CP has been proposed as an important antioxidant in reducing inflammation and acute phase response by scavenging superoxide and other reactive oxygen species [37].

Thiols contain the sulfhydryl group attached to a carbon atom. They are efficient antioxidants protecting cells against consequences of damage induced by free radicals [38,39]. In this study, levels of thiol compounds were significantly increased in patients with *P. vivax* malaria with jaundice compared with *P. vivax* malaria without jaundice. Although the thiols levels in malarial patients are not significantly higher compared to the control group, results suggest that malarial patients who developed jaundice have greater oxidative stress, and thiol compounds may be trying to restore the plasmatic balance.

Several reports in the literature suggest that drugs used to treat malaria, such as chloroquine and primaquine) lead to oxidative stress, particularly in erythrocytes [40-42]. However, in this study, patients from both groups were systematically treated with these same drugs in similar dosages, as part of the national policy, allowing therefore comparability.

Bilirubin has antioxidant properties as well as pro-oxidant. At low concentrations, it acts as a scavenger of reactive oxygen species, reducing the damage caused to the cells. However, at high concentrations, as is the case of the patients with *P. vivax* malaria who developed jaundice, bilirubin has deleterious effects on tissues. It develops oxidative stress by generating intracellular ROS in hepatic cells and cause lipid peroxidation [43]. Furthermore, bilirubin can also induce apoptosis [43], complementing the information that malaria infection induces the generation of hydroxyl radical (\bullet OH) in the liver, which may be responsible for the induction of oxidative stress and apoptosis in cells of this organ [21,22]. However, if on one side indirect bilirubin is a surrogate of haemolysis and contribute to reinforce cholestasis (jaundiced patients with lower haemoglobin levels and increase in lactate dehydrogenase support that), this compound may be faced either as a product of oxidative stress responses during malarial infection or as an inducer of oxidative stress, due to a rise in lipid and protein oxidation, ROS content, impairing glutathione metabolism (decrease of the GSH/GSSG ratio) [44]. Moreover, other studies have demonstrated that oxidative stress is increased in patients with cholecystectomy as well as in patients who developed other cholestatic diseases, and was associated with jaundice of different origin and severity [45,46].

Conclusions

In summary, the oxidative stress in *P. vivax* patients presenting jaundice is increased. Levels of oxygen reactive species may be closely linked to the damage caused by the parasite and the subsequent release of high concentrations of bilirubin in the serum. Further studies are needed to understand the mechanisms involved in liver damage in jaundiced patients, and also to validate if similar findings are seen in other less frequent complications of *P. vivax* infection, e.g., severe anaemia, coma, acute renal failure and respiratory distress. These studies may provide further evidence for better management of *P. vivax* infections and possible future anti-oxidant supportive therapy.

Competing interests

The authors declared that they have no competing interests.

Authors' contributions

CF and RCMN carried out all the biochemical analysis and drafted the manuscript, together with PL. GCM coordinated and performed all the microbiological tests. BMLM and MAAA performed the full clinical characterization of the enrolled patients. CF, MVGL and ESL participated in the design of the study. MVGL and ESL conceived of the study, and participated in its design and coordination. All authors read and approved the final manuscript.

Acknowledgements

To the patients and personnel of the *Fundação de Medicina Tropical Dr. Heitor Vieira Dourado*; and the financial support provided by CAPES, *INCT Redoxoma* and PRONEX- Malaria Network (FAPEAM/CNPq). E.S. Lima and M.V. G. Lacerda are productivity fellows level 2 from CNPq.

Author details

¹Faculty of Pharmaceutical Sciences, Universidade Federal do Amazonas, Manaus, AM 69010-300, Brazil. ²Institute of Biochemistry and Genetics, Universidade Federal de Uberlândia, Minas, MG 38400-902, Brazil. ³Fundação de Medicina Tropical Dr. Heitor Vieira Dourado, Manaus, AM 69040-000, Brazil. ⁴Universidade do Estado do Amazonas, Manaus, AM 69040-000, Brazil. ⁵Institute of Medical Virology, Charité – Universitätsmedizin Berlin, D-10117 Berlin, Germany.

Received: 18 February 2013 Accepted: 9 September 2013

Published: 10 September 2013

References

1. Gething PW, Elyazar IR, Moyes CL, Smith DL, Battle KE, Guerra CA, Patil AP, Tatem AJ, Howes RE, Myers MF, George DB, Horby P, Wertheim HF, Price RN, Mueller I, Baird JK, Hay SI: **A long neglected world malaria map: *Plasmodium vivax* endemicity in 2010.** *PLoS Negl Trop Dis* 2012, **6**:e1814.
2. Tijtra E, Anstey NM, Sugiarto P, Warikar N, Kenangalem E, Karyana M, Lampah DA, Price RN: **Multidrug-resistant *Plasmodium vivax* associated with severe and fatal malaria: a prospective study in Papua.** *Indonesia PLoS Med* 2008, **5**:e128.
3. Lomar AV, Vidal JE, Lomar FP, Barbas CV, Matos GJ, Boulos M: **Acute respiratory distress syndrome due to vivax malaria: case report and literature review.** *Braz J Infect Dis* 2005, **9**:425-430.
4. Oliveira-Ferreira J, Lacerda MVG, Brasil P, Ladislau JLB, Tauil PL, Daniel-Ribeiro CT: **Malaria in Brazil: an overview.** *Malar J* 2010, **9**:1-15.
5. Santos-Cimiera PD, Roberts DR, Alecrim MGC, Costa MR, Quinnan GV: **Malaria diagnosis and hospitalization trends.** *Emerg Infect Dis* 2007, **13**:1597-1600.
6. Ramos Junior WM, Sardinha JF, Costa MR, Santana VS, Alecrim MGC, Lacerda MV: **Clinical aspects of hemolysis in patients with *P.vivax* malaria treated with primaquine, in the Brazilian Amazon.** *Braz J Infect Dis* 2010, **14**:410-412.

7. Sarkar D, Ray S, Saha M, Chakraborty A, Talukdar A: **Clinico-laboratory profile of severe *Plasmodium vivax* malaria in a tertiary care centre in Kolkata.** *Trop Parasitol* 2013, **3**:53–57.
8. Alexandre MA, Ferreira CO, Siqueira AM, Magalhaes BL, Mourao MPG, Lacerda MVG, Alecrim MGC: **Severe *Plasmodium vivax* malaria, Brazilian Amazon.** *Emerg Infect Dis* 2010, **16**:1611–1614.
9. Kochar DK, Das A, Kochar SK, Saxena V, Sirohi P, Garg S, Kochar A, Khatri MP, Gupta V: **Severe *Plasmodium vivax* malaria: a report on serial cases from Bikaner in northwestern India.** *Am J Trop Med Hyg* 2009, **80**:194–198.
10. Kochar DK, Saxena V, Singh N, Kochar SK, Kumar SV, Das A: ***Plasmodium vivax* malaria.** *Emerg Infect Dis* 2005, **11**:132–134.
11. WHO: **Severe falciparum malaria.** *Trans R Soc Trop Med Hyg* 2000, **94**:S1–S90.
12. WHO: **Guidelines for the treatment of malaria.** Second edition. Geneva: WHO Press; 2010.
13. Anand AC, Puri P: **Jaundice in malaria.** *J Gastroenterol Hepatol* 2005, **20**:1322–1332.
14. Lanca EF, Magalhães BL, Vitor-Silva S, Siqueira AM, Benzecry SG, Alexandre MA, O'Brien C, Bassat Q, Lacerda MV: **Risk factors and characterization of *Plasmodium vivax*-associated admissions to pediatric intensive care units in the Brazilian Amazon.** *PLoS One* 2012, **7**:e35406.
15. Steinbrenner H, Sies H: **Protection against reactive oxygen species by selenoproteins.** *Biochim Biophys Acta* 2009, **1790**:1478–1485.
16. Araujo CF, Lacerda MVG, Abdalla DSP, Lima ES: **The role of platelet and plasma markers of antioxidant status and oxidative stress in thrombocytopenia among patients with vivax malaria.** *Mem Inst Oswaldo Cruz* 2008, **103**:517–521.
17. Sohail M, Kaul A, Raziuddin M, Adak T: **Decreased glutathione-S-transferase activity: diagnostic and protective role in vivax malaria.** *Clin Biochem* 2007, **40**:377–382.
18. Narsaria N, Mohanty C, Das BK, Mishra SP, Praosad R: **Oxidative stress in children with severe malaria.** *J Trop Pediatr* 2011, **58**:1–4.
19. Kapoor G, Banyal HS: **Glutathione reductase and thioredoxin reductase: novel antioxidant enzymes from *Plasmodium berghei*.** *Korean J Parasitol* 2009, **47**:421–424.
20. Kawazu S, Yasuda-Komaki K, Oku H, Kano S: **Peroxiredoxins in malaria parasites: parasitologic aspects.** *Parasitol Int* 2008, **57**:1–7.
21. Dey S, Guha M, Alam A, Goyal M, Bindu S, Pal C, Maity P, Mitra K, Bandyopadhyay U: **Malarial infection develops mitochondrial pathology and mitochondrial oxidative stress to promote hepatocyte apoptosis.** *Free Radic Biol Med* 2009, **46**:271–281.
22. Guha M, Kumar S, Choubey V, Maity P, Bandyopadhyay U: **Apoptosis in liver during malaria: role of oxidative stress and implication of mitochondrial pathway.** *FASEB J* 2006, **20**:339–449.
23. Brewer GJ, Tarlov AR, Alving AS: **Methaemoglobin reduction test.** *Bull World Health Organ* 1960, **22**:633–640.
24. Snounou G, Viriyakosol S, Zhu XP, Jarra W, Pinheiro L, Rosario VE, Thaithong S, Brown KN: **High sensitivity of detection of human malaria parasites by the use of nested polymerase chain reaction.** *Mol Biochem Parasitol* 1993, **61**:315–320.
25. Esterbauer H, Cheeseman KH: **Determination of aldehydic lipid peroxidation products: malonaldehyde and 4-hydroxynonenal.** *Methods Enzymol* 1990, **186**:407–421.
26. Holmgren A, Bjornstedt M: **Thioredoxin and thioredoxin reductase.** *Methods Enzymol* 1995, **252**:199–208.
27. Schosinsky KH, Lehmann HP, Beeler MF: **Measurement of ceruloplasmin from its oxidase activity in serum by use of o-dianisidine dihydrochloride.** *Clin Chem* 1974, **20**:1556–1563.
28. Ellman GL: **Tissue sulfhydryl groups.** *Arch Biochem Biophys* 1959, **82**:70–77.
29. Hu ML: **Measurement of protein thiol groups and glutathione in plasma.** *Methods Enzymol* 1994, **233**:380–385.
30. Lacerda MV, Fragoso SC, Alecrim MG, Alexandre MA, Magalhaes BM, Siqueira AM, Ferreira LC, Araujo JR, Mourao MP, Ferrer M, Castillo P, Martin-Jaular L, Fernandez-Becerra C, Del Portillo H, Ordi J, Alonso PL, Bassat Q: **Postmortem characterization of patients with clinical diagnosis of *Plasmodium vivax* Malaria: to what extent does this parasite kill?** *Clin Infect Dis* 2012, **55**:e67–e74.
31. Pabón A, Carmona J, Burgos LC, Blair S: **Oxidative stress in patients with non-complicated malaria.** *Clin Biochem* 2003, **36**:71–78.
32. Sohail M, Kumar R, Kaul A, Arif E, Kumar S, Adak T: **Polymorphism in glutathione S-transferase P1 is associated with susceptibility to *Plasmodium vivax* compared to *P. falciparum* and upregulates the GST level during malarial infection.** *Free Radic Biol Med* 2010, **49**:1746–1754.
33. Nordberg J, Arnér ESJ: **Reactive oxygen species, antioxidants, and the mammalian thioredoxin system.** *Free Radic Biol Med* 2001, **31**:1287–1312.
34. Kim S, Jung H, Hyum D, Park E, Kim Y, Lim C: **Glutathione reductase plays an anti-apoptotic role against oxidative stress in human hepatoma cells.** *Biochimie* 2010, **92**:927–932.
35. Erel O, Kocyigit A, Avci S, Aktepe N, Bulut V: **Oxidative Stress and antioxidative status of plasma and erythrocytes in patients with vivax malaria.** *Clin Biochem* 1997, **30**:631–639.
36. Das BS, Patnaik JK, Mohanty S, Satpathy SK: **Plasma antioxidants and lipid peroxidation products in falciparum malaria.** *Am J Trop Med Hyg* 1993, **49**:720–725.
37. Ehrenwald E, Chisolm GM, Fox PL: **Intact human ceruloplasmin oxidatively modifies low density lipoprotein.** *J Clin Invest* 1994, **93**:1493–1501.
38. Costa CM, Dos Santos RCC, Lima ES: **A simple automated procedure for thiol measurement in human serum samples.** *J Bras Patol Med Lab* 2006, **42**:345–350.
39. Hansen RE, Winther JR: **An introduction to methods for analyzing thiols and disulfides: reactions, reagents, and practical considerations.** *Anal Biochem* 2009, **394**:147–158.
40. Baird JK, Hoffman SL: **Primaquine therapy for malaria.** *Clin Infect Dis* 2004, **39**:1336–1345.
41. Bolchoz LJC, Morrow DJ, Jollow DJ, Mcmillan DC: **Primaquine-induced hemolytic anemia: effect of 6-methoxy-8-hydroxyaminoquinoline on rat erythrocyte sulfhydryl status, membrane lipids, cytoskeletal proteins, and morphology.** *J Pharmacol Exp Ther* 2002, **303**:141–148.
42. Becker K, Tilley L, Vennerstrom JL, Roberts D, Rogerson S, Ginsburg H: **Oxidative stress in malaria parasite-infected erythrocytes: host-parasite interactions.** *Int J Parasitol* 2004, **34**:163–189.
43. Kumar S, Guha M, Choubey V, Maity P, Srivastava K, Puri SK, Bandyopadhyay U: **Bilirubin inhibits *Plasmodium falciparum* growth through the generation of reactive oxygen species.** *Free Radic Biol Med* 2008, **44**:602–613.
44. Brito MA, Brites D, Butterfield DA: **A link between hyperbilirubinemia, oxidative stress and injury to neocortical synaptosomes.** *Brain Res* 2004, **1026**:33–43.
45. Miranda-Díaz AG, Hermosillo-Sandoval JM, Ortiz GG, Lizardi-García D: **Serum oxidative stress is increased in patients with post cholecystectomy bile duct injury.** *Rev Esp Enferm Dig* 2010, **102**:353–356.
46. Silina EV, Stupin VA, Garkhranov TV, Khokonov MA, Bolevich SB, Men'shova NI, Sineĭnikova TG: **Oxidative stress in patients with mechanical jaundice of different origin and severity.** *Klin Med* 2011, **89**:57–63.

doi:10.1186/1475-2875-12-315

Cite this article as: Fabbri et al.: Lipid peroxidation and antioxidant enzymes activity in *Plasmodium vivax* malaria patients evolving with cholestatic jaundice. *Malaria Journal* 2013 **12**:315.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

