Malaria Journal



Review Open Access

The role of anti-malarial drugs in eliminating malaria Nicholas I White^{1,2}

Address: ¹Mahidol – Oxford Tropical Medicine Research Unit, Faculty of Tropical Medicine, Mahidol University, 420/6 Rajvithi Road, Bangkok 10400, Thailand and ²Centre for Clinical Vaccinology and Tropical Medicine, Churchill Hospital, Oxford, UK

Email: Nicholas J White - nickw@tropmedres.ac

Published: 11 December 2008

Malaria Journal 2008, **7**(Suppl 1):S8 doi:10.1186/1475-2875-7-S1-S8

This article is available from: http://www.malariajournal.com/content/7/S1/S8

© 2008 White; licensee BioMed Central Ltd.

This is an open access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/2.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Effective anti-malarial drug treatment reduces malaria transmission. This alone can reduce the incidence and prevalence of malaria, although the effects are greater in areas of low transmission where a greater proportion of the infectious reservoir is symptomatic and receives anti-malarial treatment. Effective treatment has greater effects on the transmission of falciparum malaria, where gametocytogenesis is delayed, compared with the other human malarias in which peak gametocytaemia and transmissibility coincides with peak asexual parasite densities. Mature Plasmodium falciparum gametocytes are more drug resistant and affected only by artemisinins and 8-aminoquinolines. The key operational question now is whether primaquine should be added to artemisinin combination treatments for the treatment of falciparum malaria to reduce further the transmissibility of the treated infection. Radical treatment with primaquine plays a key role in the eradication of vivax and ovale malaria. More evidence is needed on the safety of primaquine when administered without screening for G6PD deficiency to inform individual and mass treatment approaches in the context of malaria elimination programmes.

Background

Anti-malarial drugs play a central role in the control and ultimate elimination of malaria, but, in most circumstances, they cannot do the job alone. Whether malaria can be eliminated with current tools is a subject of active debate [1], but it is generally agreed that the tools to reduce substantially the global burden of malaria are available. The corollary is that if the current classes of effective anti-malarial drugs (notably the artemisinin derivatives) are lost, then effective control and elimination will not be possible. Anti-malarials, if effective, reduce the transmission of malaria, but the relationship between efficacy and transmission reduction is not straightforward. This relationship is the subject of this review.

Epidemiological considerations

Where transmission of malaria is intense, even highly effective interventions which reduce mortality may have little noticeable effect on the clinical pattern of malaria. A 95% reduction in transmission from 500 infective bites per year to 25 will not change the incidence or prevalence of malaria noticeably. In this context, there is a tremendous redundancy in the transmission reservoir (from asymptomatic gametocyte carriers), and the use of effective anti-malarial drugs will have relatively little impact on transmission intensity. For example, in a high transmission setting in Western Kenya, it was estimated, based on mosquito feeding studies, that 28% of all transmission derived from children ≥10 years and adults - a largely asymptomatic population [2]. This would correspond to 10-50 infectious bites per year, plenty to sustain a hyper or holoendemic pattern of malaria even if every symptomatic patient received highly effective drugs. But as transmission intensity falls, the premunition of the population declines, and an increasing proportion of transmission derives from symptomatic individuals, who will seek antimalarial treatment. In a detailed epidemiological study conducted in a low transmission setting on the Thai-Burmese border (entomological inoculation rate <1 per species) all P. falciparum infections were symptomatic, but approximately 10% of *Plasmodium vivax* infections were still asymptomatic [3]. Although this is probably representative of most low transmission settings, it is important to note that transmission intensity is remarkably heterogeneous over short geographic distances, and that low transmission areas usually contain small foci of much higher transmission intensity which act as reservoirs of infection. Inhabitants of these small foci sustain malaria over the annual dry season. Premunition does not change abruptly, so if there are sudden decreases (or increases) in transmission intensity then the changes in clinical epidemiology lag behind (hysteresis). Thus, the effects of antimalarial treatment on the incidence and prevalence of malaria increase as transmission falls. Unfortunately, there are no precise estimates of this relationship.

Biological considerations

As human malaria is transmitted by sexual stages of the parasites, not asexual stages, infecting anopheline mosquito vectors, transmission depends upon the duration for which gametocytes are carried in the blood, the infectivity of this gametocytaemia to the local vectors, and the abundance and behaviour of the vectors [4]. The attractiveness of humans to mosquitoes is unevenly distributed amongst the human population. Mosquitoes apparently prefer to bite individuals with lower biomass and lower body surface area [5], pregnant women [6], and those with smelly feet [7,8], amongst other things. Gametocytaemia is detectable by microscopy down to densities of approximately 10-20/uL. At least one male gametocyte's progeny (eight microgametes) and one female macrogamete are required in a mosquito blood meal (approx 2-3 μ L) for infection to occur. Thus, gametocyte densities of 1/ μL can theoretically infect vector mosquitoes [4,9]. This is below the density which can be detected by routine microscopy. It has sometimes been stated that malaria can be transmitted (to mosquitoes) by individuals who are not carrying gametocytes. This is impossible. What is actually meant is that the individuals obviously were carrying viable gametocytes in their blood, but the density was below that detectable by microscopy [10]. As the progeny of at least two gametes are required to form a zygote, the probabilities of transmission as gametocyte densities fall below 1/µL rapidly become vanishingly small. Whether gametocytes concentrate in the dermal capillaries (as they ought to!) still remains unclear, but even if they do, these numerical considerations still apply. Plasmodium falciparum differs from the other three human malarias in two important respects; first, gametocyte formation is delayed with respect to the peak production of asexual stages [11-13], and, second, that mature gametocytes are resistant to most of the anti-malarial drugs, which affect asexual stages. The developing sexual stages of P. falciparum (stages I to IV) remain sequestered in the microvasculature for approximately 10 days before appearing as morphologically distinct male and female gametocytes (stage 5) in the peripheral blood. Peak transmissibility coincides approximately with peak gametocytaemia [12]. Several factors increase the production of gametocytes in falciparum malaria, most of which can be considered under the general rubric of "stress"; these include long duration of infection, anaemia, partially effective immune responses, and partially effective drugs [14-17].

In infections with *P. vivax, P. malariae* and *P. ovale,* the asexual and sexual stages appear almost together [18-20], and, in contrast to *P. falciparum* [19], are sensitive to all the drugs which kill the asexual stages [21] These three malaria parasites transmit very efficiently at low parasite densities – much more efficiently than *P. falciparum.* Thus, a greater proportion of infected subjects without detectable gametocytaemia can transmit these malaria infections. Because the majority of patients presenting with these so-called benign malarias are already infective to biting anopheline mosquito vectors when they present ill for treatment, administration of effective anti-malarial drugs has less effect on overall transmission compared with falciparum malaria.

Pharmacological considerations

All anti-malarial drugs which kill asexual stages also kill the early stages of P. falciparum gametocytes, as well as the mature P. vivax, malariae, and ovale gametocytes. Thus the combination of reducing the asexual stage progenitors of the sexual stages, and killing the gametocytes themselves reduces transmissibility. In general, the asexual and sexual stage activities parallel each other [21-24], although the quantitative relationships between anti-malarial drug concentrations (pharmacokinetics; PK) and reduction in transmissibility (pharmacodynamics; PD) are not well characterized [25], particularly for vivax, malariae, and ovale infections. For fully sensitive asexual stage parasites, the artemisinin derivatives produce the most rapid reductions in parasitaemia [26,27], followed by chloroquine [28], then closely behind the other quinolines and related drugs, together with atovaquone, and finally the antifols. It is not clear whether amodiaquine, piperaquine and pyronaridine are closer to chloroquine than quinine and mefloquine in this batting order but if they are, the differences are small. The antibiotics with anti-malarial activity are generally less active and clear parasitaemia more slowly than the anti-malarial drugs [23]. At a population

level there is wide inter-individual variation in the effects of a single anti-malarial treatment on the reduction in parasitaemia. This derives from host differences in anti-malarial adherence, absorption, distribution (drug elimination is less relevant), and splenic function as well as differences in the numbers, stage, synchronicity and susceptibility of the infecting parasites [26]. Anti-malarial drug susceptibility of the infecting parasites becomes important when the drug concentrations in blood do not produce the maximum effect (i.e. are below the minimum parasiticidal concentration; MPC). The proportion of treated patients with blood concentrations below the MPC is a function of drug quality, dose taken, host pharmacokinetics, and resistance. As resistance worsens this proportion increases [29].

The overall effects of anti-malarial drugs on P. falciparum gametocytaemia (i.e. late stage gametocytes which circulate and are detectable by microscopy) is a composite of their effects on the progenitors, the early (more drug susceptible) stage 1 to 3 sequestered gametocytes, and the more mature (less drug susceptible) stages. P. falciparum gametocytaemia following antifol treatment (best documented for sulphadoxine-pyrimethamine) is consistently greater than that following other drug classes [4,30,31], whereas treatment with the artemisinin derivatives is associated with lower rates of gametocyte carriage [13,14,27,32,33]. The 8-aminoquinolines (primaquine, quinocide, tafenoquine) occupy a unique position. Most data refer to primaquine, the only generally available compound. Primaquine has reasonable asexual stage activity against *P. vivax* malaria [34], kill hypnozoites of *P.* vivax and P.ovale [35,36], and mature gametocytes of P. falciparum [37,38], but importantly lacks useful activity against asexual stages (and presumably early gametocytes) of P. falciparum [37,38]. It is not known how primaquine works in any of these three actions, nor which of its many metabolites is the active moiety. The considerable variations in immunity, pharmacokinetics and pharmacodynamics results in marked variability transmissibility related to the activity of anti-malarial drugs [39-43].

Anti-malarial drug resistance and transmissibility

In considering anti-malarial drug effects on transmissibility, three different components need to be considered; a) activity against asexual stages and early gametocytes, b) activity against mature infectious gametocytes, and c) sporontocidal effects in the mosquito. As drug activity against the asexual stage activity falls because of worsening antimalarial resistance, the rate at which parasitaemia is reduced falls, and treatment failure rates increase. Gametocytaemia is a very sensitive measure of worsening drug resistance. As SP resistance worsened in South Africa, increasing gametocytaemia was the first warning sign.

Importantly it preceded measurable changes in parasite clearance or a decline in cure rates [44]. Thus, transmission, and in particular transmission of resistance, would have increased before detectable changes in treatment failure rates. Okell et al have recently pooled data from 3,174 patients enrolled in six anti-malarial trials conducted in The Gambia and Kenya. ACT treatment (either artesunate-SP, artesunate-chloroquine or artemether lumefantrine) was associated with a significant reduction in the probability of being gametocytaemic on the day of transmission experiments (OR 0.20 95% CI 0.16-0.26), transmission to mosquitoes by slide-positive gametocyte carriers (OR mosquito infection 0.49 95% CI 0.33-0.73), and AUC of gametocyte density (ratio of means 0.35 95% CI 0.31-0.41) [45]. Partially effective anti-malarial drug treatment increases gametocyte carriage both by reducing asexual parasite killing and by providing a stress on the surviving asexual parasite population [16,17]. These factors allow a greater fraction of asexual parasites to switch to gametocyte development. The chance of an infection recrudescing also increases. Recrudescent infections are more likely than primary infections to be gametocytaemic at presentation (because of the extended duration of infection, drug "stress" and anaemia), and are also more likely to fail subsequent treatment. The net result of the increased gametocyte carriage of the primary infections, and any subsequent recrudescences, is much greater gametocyte carriage in infections caused by resistant parasites. This translates (with considerable inter-individual variation) into increased transmissibility. It is this transmission advantage that drives the spread of resistance.

For antifols and atovaquone, there are also effects in the mosquito to consider. These drugs prevent the formation of sporozoites by interfering with oocyst development in the anopheline mosquito [46,47]. Tafenoquine also has sporontocidal activity and is more active than primaquine which has little or no activity [48]. Antifol resistant parasites are more transmissible than antifol sensitive parasites in the presence of drug indicating a further location of selective pressure driving the spread of antifol resistance [49].

It is evident that anti-malarial drug resistance spreads because of the greater transmission potential of the resistant parasites in the presence of the anti-malarial drug. Most currently available anti-malarial drugs are eliminated slowly from the body and so, if they are used intensely in a malaria endemic area, a significant proportion of the community has variable concentrations of the drug in their blood. These concentrations act as a selective filter favouring the establishment of resistant infections. The degree of selection depends on several factors including the intensity of transmission, the immunity profile of

the host and the pharmacokinetic and pharmacodynamic properties of the anti-malarial drug.

Benefits of reducing transmission

Intense malaria is associated with a considerable burden of morbidity and mortality in childhood. Infant mortality rates are positively correlated with transmission intensity measured by the EIR (estimated entomological inoculation rate), although the exact relationship is not well defined [50]. Mosquitoes may bite infants proportionally more than adults; they have been shown to bite preferentially individuals with lower biomass and lower body surface [5,50]. Deployment of effective anti-malarial drugs either alone or in combination with other control measures, particularly insecticide treated bed nets and indoor residual spraying, reduces malaria morbidity and mortality [51-53]. Indeed, the introduction of effective malaria control reduces the mortality rate more than would be expected by the direct prevention of the malaria deaths alone. This is explained by many of the conditions leading to infant deaths being only indirectly or cumulatively related to malaria, such as malaria anaemia and low birth weight due to placental malaria. Importantly reducing infant mortality does not seem to be balanced by higher mortality later on [54]. Thus, there is overwhelming evidence for a benefit from reducing malaria transmission.

Operational considerations

Artemisinin combination treatments are the recommended first-line drugs for the treatment of falciparum malaria in endemic areas. They are highly effective and well tolerated, but despite considerable increases in recent support for anti-malarial drugs, the majority of those that need ACTs do not receive them. Scaling up deployment of ACTs and insecticide-treated nets, and importantly subsidizing ACTs so that the treatment is affordable, and donating insecticide-treated nets, will have a major impact on malaria morbidity and mortality. Indoor residual spraying also has a major role to play in some areas. Should elimination programmes do anything different? The outstanding unresolved question for treatment, is whether a single "gametocytocidal" dose of primaquine (0.5 to 0.75 mg base/kg) should be added to ACT for falciparum malaria treatment [55]. Addition of a single dose of primaquine to first-line treatment has been recommended by malaria control programmes in some areas, but not others, for many years. Despite this there is remarkably little evidence on either safety or efficacy. Prevention of relapse in vivax and ovale infections, and thus control of transmission, depends on taking a two-week radical curative regimen of primaguine (i.e. 14 days of 0.25 to 0.5 mg base/kg/day). Again this is recommended in some areas, and not in others.

Although uncertain efficacy, and the difficulty in ensuring adherence to 14-day regimens, have lowered enthusiasm for pursuing radical cure in endemic areas, the main reservations which have limited the use of primaquine concern safety. Primaquine predictably causes abdominal discomfort. The effect is dose-dependent, but considerably ameliorated by food. The 8-aminoquinolines are oxidant drugs and cause oxidant haemolysis. Methaemoglobinaemia is usual [56], but in inherited red cell enzyme deficiencies which impair defences against oxidant stress, haemolysis can be severe and occasionally life-threatening [57]. The introduction of primaquine in 1951 led to the discovery of glucose-6-phosphate dehydrogenase (G6PD) deficiency, the most common human enzyme deficiency, which affects approximately 400 million people worldwide [58]. There are some 140 different genotypes of G6PD deficiency [59,60] with corresponding phenoytypes that vary from mild to severe. The extent of oxidant haemolysis depends on the degree and nature of deficiency. Enzyme deficiency protects against severe malaria and so the abnormality is prevalent mainly in areas where malaria is or was prevalent. It is often recommended that G6PD deficiency testing be performed before administering treatment. In reality testing is usually unavailable. Several rapid tests are available in theory (but deployment is very limited) and there is insufficient evidence on their sensitivity and specificity to recommend their general deployment. The semi-quantitative fluorescence spot test is more demanding but reliable, and quantitation requires a spectrophotometric assay. So, in practice testing is usually unavailable and primaquine is often not given.

Mass treatment

Mass treatment of the entire population with anti-malarial drugs was performed on many occasions in the eradication programmes of the 1950s and 1960s, and has been employed sporadically since then. This experience has been reviewed recently [61]. As a strategy it has not found favour in recent years. The alternative proactive approaches are mass screening and treatment (MST). This has been preferred over mass treatment in Western Cambodia in current efforts to contain and eliminate malaria there. MST is a more logistically demanding approach, and it assumes that most of the infectious reservoir will have detectable parasitaemias at the time of screening. Obviously, for P. vivax and P. ovale, screening will not detect hypnozoite carriage. The advantages and disadvantages of these two different approaches bear further examination and evaluation. Mass treatment and MST would be more acceptable if toxicity concerns over administration of the 8 aminoquinolines, without screening for G6PD deficiency, in the population can be overcome.

Competing interests

The author is co-chairman of the WHO GMP technical expert group on the prevention and treatment of malaria.

Acknowledgements

This article has been published as part of Malaria Journal Volume 7 Supplement 1, 2008: Towards a research agenda for global malaria elimination. The full contents of the supplement are available online at http:// www.malariajournal.com/supplements/7/SI

References

- Mendis K, Rietveld A, Warsame M, Bosman A, Greenwood B, Wernsdorfer W: From malaria control to eradication: The WHO perspective. Lancet in press.
- Githeko AK, Brandling-Bennett AD, Beier M, Atieli F, Owaga M, Collins FH: The reservoir of Plasmodium falciparum malaria in a holoendemic area of western Kenya. Trans R Soc Trop Med Hyg 1992, 86:355-358.
- Luxemburger C, Kyaw Ley Thew, White NJ, Webster HK, Kyle DE, Maelankiri L, Chongsuphajaisiddhi T, Nosten F: The epidemiology of malaria in a Karen population on the western border of Thailand. Trans R Soc Trop Med Hyg 1996, 90:105-111
- Drakeley C, Sutherland C, Bousema JT, Sauerwein RW, Targett GA: The epidemiology of Plasmodium falciparum gametocytes: weapons of mass dispersion. Trends Parasitol 2006, 22:424-430. Port GR, Boreham PFL, Bryan JH: The relationship of host size to
- feeding by mosquitoes of the Anopheles gambiae Giles complex (Diptera, Culicidae). Bull Entomol Res 1980, 70:133-144.
- Ansell J, Hamilton KA, Pinder M, Walraven GE, Lindsay SW: Shortrange attractiveness of pregnant women to Anopheles gambiae mosquitoes. Trans R Soc Trop Med Hyg 2002, 96:113-116.
- Knols BG, De Jong R: Limburger cheese as an attractant for the malaria mosquito. Anopheles gambiae s.s. Parasitol Today 1996, 12:159-161
- Murphy MW, Dunton RF, Perich MJ, Rowley WA: Attraction of Anopheles (Diptera: culicidae) to volatile chemicals in Western Kenya.] Med Entomol 2001, 38:242-244.
- Jeffery GM, Eyles DE: Infectivity to mosquitoes of Plasmodium falciparum as related to gametocyte density and duration of infection. Am J Trop Med Hyg 1955, 4:781-789.
- 10. Schneider P, Bousema JT, Gouagna LC, Otieno S, Vegte-Bolmer M van de, Omar SA, Sauerwein RW: Submicroscopic Plasmodium falciparum gametocyte densities frequently result in mosquito infection. Am J Trop Med Hyg 2007, 76:470-474.
- Eichner M, Diebner HH, Molineaux L, Collins WE, Jeffery GM, Dietz K: Genesis, sequestration and survival of Plasmodium falciparum gametocytes: parameter estimates from fitting a model to malariatherapy data. Trans R Soc Trop Med Hyg 2001, 95:497-501.
- 12. Collins WE, Jeffery GM: A retrospective examination of mosquito infection on humans infected with Plasmodium falciparum. Am J Trop Med Hyg 2003, 68:366-371.
- McKenzie FE, Jeffery GM, Collins WE: Gametocytemia and fever in human malaria infections. J Parasitol 2007, 93:627-633.
- Bruce MC, Alano P, Duthie S, Carter R: Commitment of the malaria parasite Plasmodium falciparum to sexual and asexual development. Parasitology 1990, 100(Pt 2):191-200.
- Smith TG, Lourenço P, Carter R, Walliker D, Ranford-Cartwright LC: Commitment to sexual differentiation in the human malaria parasite, Plasmodium falciparum. Parasitology 2000, 121:127-133.
- Price RN, Nosten F, Luxemburger C, Phaipun L, ter Kuile F, van Vugt M, Chongsuphajaisiddhi T, White NJ: Risk factors for gametocyte carriage in uncomplicated falciparum malaria. Am J Trop Med Hyg 1999, 60:1019-1023.
- 17. von Seidlein L, Drakeley C, Greenwood B, Walraven G, Targett G: Risk factors for gametocyte carriage in Gambian children. Am J Trop Med Hyg 2001, 65:523-527.
- 18. McKenzie FE, Jeffery GM, Collins WE: Plasmodium vivax bloodstage dynamics. | Parasitol 2002, 88:521-535
- McKenzie FE, Jeffery GM, Collins WE: Plasmodium malariae bloodstage dynamics. J Parasitol 2001, 87:626-637.
- Collins WE, Jeffery GM: A retrospective examination of sporozoite-induced and trophozoite-induced infections with Plas-

- modium ovale: development of parasitologic and clinical immunity during primary infection. Am | Trop Med Hyg 2002, **66:**492-502.
- Pukrittayakamee S, Imwong M, Stepniewska K, Day NPJ, White NJ: The effects of different antimalarial drugs on gametocyte carriage in P. vivax malaria. Am J Trop Med Hyg 2008, **79**:378-384. 22. Pukrittayakamee S, Chotivanich K, Chantra A, Clemens R, Looareesu-
- wan S, White NJ: Activities of artesunate and primaquine against asexual- and sexual-stage parasites in falciparum malaria. Antimicrob Agents Chemother 2004, 48:1329-1334.
- 23. Pukrittayakamee S, Imwong M, Looareesuwan S, White NJ: Therapeutic responses to antimalarial and antibacterial drugs in vivax malaria. Acta Trop 2004, 89:351-356.
- Pukrittayakamee S, Chantra A, Simpson JA, Vanijanonta S, Clemens R, Looareesuwan S, White NJ: Therapeutic responses to different antimalarial drugs in vivax malaria. Antimicrob Agents Chemother 2000, 44:1680-1685.
- Barnes KI, White NJ: Population biology and antimalarial resistance: The transmission of antimalarial drug resistance in Plasmodium falciparum. Acta Trop 2005, 94:230-240.
- White NJ: Assessment of the pharmacodynamic properties of the antimalarial drugs in-vivo. Antimicrob Agents Chemother 1997, 41:1413-1422.
- International Artemisinin Study Group: Artesunate combinations for treatment of malaria: meta- analysis. Lancet 2004. 363:9-17
- White NJ, Krishna S, Waller D, Craddock C, Kwiatkowski D, Brewster D: Open comparison of intramuscular chloroquine and quinine in children with severe chloroquine-sensitive falciparum malaria. Lancet 1989, 2:1313-1316.
- White NJ: Antimalarial drug resistance. J Clin Invest 2004, 113:1084-92
- Butcher GA: Antimalarial drugs and the mosquito transmis-
- sion of Plasmodium. Int J Parasitol 1997, 27:975-987.
 Govere JM, Durrheim DN, Mngomezulu NM, Barnes K, Sharp B: Infectivity of Plasmodium falciparum gametocytes to Anopheles arabiensis after treatment with sulfadoxine-pyrimethamine. Trans R Soc Trop Med Hyg 2003, 97:707-708.
- Price RN, Nosten F, Luxemburger C, ter Kuile F, Paiphun L, Chongsuphajaisiddhi T, White NJ: The effects of artemisinin derivatives on malaria transmissability. Lancet 1996, 347:1654-1658.
- Hallett RL, Sutherland CJ, Alexander N, Ord R, Jawara M, Drakeley CJ, Pinder M, Walraven G, Targett GA, Alloueche A: **Combination** therapy counteracts the enhanced transmission of drugresistant malaria parasites to mosquitoes. Antimicrob Agents Chemother 2004, 48:3940-3943.
- Pukrittayakamee S, Vanijanonta S, Chantra A, Clemens R, White NJ: Blood stage antimalarial efficacy of primaquine in Plasmodium vivax malaria. J Infect Dis 1994, 169:932-935.
- Krotoski WA, Garnham PC, Cogswell FB, Collins WE, Bray RS, Gwasz RW, Killick-Kendrick R, Wolf RH, Sinden R, Hollingdale M: Observations on early and late post-sporozoite tissue stages in primate malaria. IV. Pre-erythrocytic schizonts and/or hypnozoites of Chesson and North Korean strains of Plasmodium vivax in the chimpanzee. Am J Trop Med Hyg 1986, **35:**263-274.
- Udomsangpetch R, Kaneko O, Chotivanich K, Sattabongkot J: Cultivation of Plasmodium vivax. Trends Parasitol 2008, 24:85-88.
- Arnold J, Alving AS, Hockwald RS, Clayman CB, Dern RJ, Beutler E, Flanagan CL, Jeffery GM: The antimalarial action of primaquine against the blood and tissue stages of falciparum malaria (Panama, P-F-6 strain). J Lab Clin Med 1955, 46:391-7.
- Pukrittayakamee S, Chotivanich K, Chantra A, Clemens R, Looareesuwan S, White NJ: Activities of artesunate and primaquine against asexual- and sexual-stage parasites in falciparum malaria. Antimicrob Agents Chemother 2004, 48:1329-1334.
- Burgess RW, Bray RS: The effect of a single dose of primaquine on the gametocytes, gametogony and sporogony of Laverania falciparum. Bull World Health Organ 1961, 24:451-456.
- Tchuinkam T, Mulder B, Dechering K, Stoffels H, Verhave JP, Cot M, Carnevale P, Meuwissen JH, Robert V: Experimental infections of Anopheles gambiae with Plasmodium falciparum of naturally infected gametocyte carriers in Cameroon: factors influencing the infectivity to mosquitoes. Trop Med Parasitol 1993, 44:271-276

- Coleman RE, Nath AK, Schneider I, Song G-H, Klein TA, Milhous WK: Transmission-blocking antimalarials prevent sporogony of Plasmodium falciparum and P. berghei in Anopheles stephensi mosquitoes. Am J Trop Med Hyg 1994, 50:646-653.
- 42. Drakeley CJ, Secka I, Correa S, Greenwood BM, Targett GA: Host haematological factors influencing the transmission of Plasmodium falciparum gametocytes to Anopheles gambiae s.s. mosquitoes. Trop Med Int Health 1999, 4:131-138.
- Hallett RL, Dunyo S, Ord R, Jawara M, Pinder M, Randall A, Alloueche A, Walraven G, Targett GA, Alexander N, Sutherland CJ: Chloroquine/sulphadoxine-pyrimethamine for Gambian children with malaria: transmission to mosquitoes of multidrugresistant Plasmodium falciparum. PLoS Clin Trials 2006, 1:e15.
- 44. Barnes KI, Little F, Mabuza A, Mngomezulu N, Govere J, Durrheim D, Roper C, Watkins B, White NJ: Increased gametocytemia after treatment: an early parasitological indicator of emerging sulfadoxine-pyrimethamine resistance in falciparum malaria. J Infect Dis 2008, 197:1605-1613.
- 45. Okell LC, Drakeley CJ, Ghani AC, Bousema T, Sutherland CJ: Reduction of transmission from malaria patients by artemisinin combination therapies: a pooled analysis of six randomized trials. Malar J 2008, 7:125.
- Teklehaimanot A, Nguyen-Dinh P, Collins WE, Barber AM, Campbell CC: Evaluation of sporontocidal compounds using Plasmodium falciparum gametocytes produced in vitro. Am J Trop Med Hyg 1985, 34:429-34.
- Ponsa N, Sattabongkot J, Kittayapong P, Eikarat N, Coleman RE: Transmission-blocking activity of tafenoquine (WR-238605) and artelinic acid against naturally circulating strains of Plasmodium vivax in Thailand. Am J Trop Med Hyg 2003, 69:542-547.
- Omar MS, Collins WE, Contacos PG: Gametocytocidal and sporontocidal effects of antimalarial drugs on malaria parasites.
 II. Action of the folic reductase inhibitors, chlorguanide, and pyrimethamine against Plasmodium cynomolgi. Exp Parasitol 1974, 36:167-77.
- Méndez F, Herrera S, Murrain B, Gutiérrez A, Moreno LA, Manzano M, Muñoz A, Plowe CV: Selection of antifolate-resistant Plasmo-dium falciparum by sulfadoxine-pyrimethamine treatment and infectivity to Anopheles mosquitoes. Am J Trop Med Hyg 2007. 77:438-443.
- Smith T, Killeen G, Lengeler C, Tanner M: Relationships between the outcome of *Plasmodium falciparum* infection and the intensity of transmission in Africa. Am J Trop Med Hyg 2004, 71(2 Suppl):80-86.
- Barnes KI, Durrheim DN, Little F, Jackson A, Mehta U, Allen E, Dlamini SS, Tsoka J, Bredenkamp B, Mthembu DJ, White NJ, Sharp BL: Effect of artemether-lumefantrine policy and improved vector control on malaria burden in KwaZulu-Natal, South Africa. PLoS Med 2005, 2:e330.
- Bhattarai A, Ali AS, Kachur SP, Mårtensson A, Abbas AK, Khatib R, Al-Mafazy AW, Ramsan M, Rotllant G, Gerstenmaier JF, Molteni F, Abdulla S, Montgomery SM, Kaneko A, Björkman A: Impact of artemisinin-based combination therapy and insecticidetreated nets on malaria burden in Zanzibar. PLoS Med 2007, 4:e309.
- Carrara VI, Sirilak S, Thonglairuam J, Rojanawatsirivet C, Proux S, Gilbos V, Brockman A, Ashley EA, McGready R, Krudsood S, Leemingsawat S, Looareesuwan S, Singhasivanon P, White N, Nosten F: Deployment of early diagnosis and mefloquine-artesunate treatment of falciparum malaria in Thailand: the Tak Malaria Initiative. PLoS Med 2006, 3:e183.
- 54. Binka FN, Hodgson A, Adjuik M, Smith T: Mortality in a sevenand-a-half-year follow-up of a trial of insecticide-treated mosquito nets in Ghana. Trans R Soc Trop Med Hyg 2002, 96:597-599.
- Chomcharn Y, Surathin K, Bunnag D, Sucharit S, Harinasuta T: Effect
 of a single dose of primaquine on a Thai strain of Plasmodium
 falciparum. Southeast Asian J Trop Med Public Health 1980,
 11-408-412
- Coleman MD, Coleman NA: Drug-induced methaemoglobinaemia. Treatment issues. Drug Saf 1996, 14:394-405.
- Beutler E, Duparc S: G6PD Deficiency Working Group. Glucose-6-phosphate dehydrogenase deficiency and antimalarial drug development. Am J Trop Med Hyg 2007, 77:779-789.

- Alving AS, Carson PE, Flanagan CL, Ickes CE: Enzymatic deficiency in primaquine-sensitive erythrocytes. Science 1956, 124:484-485.
- Beutler E: G6PD deficiency. Blood 1994, 84:3613-3636.
- Beutler E: Glucose-6-phosphate dehydrogenase deficiency: a historical perspective. Blood 2008, 111:16-24.
- von Seidlein L, Greenwood BM: Mass administrations of antimalarial drugs. Trends Parasitol 2003, 19:452-460.

Publish with **Bio Med Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours you keep the copyright

Submit your manuscript here: http://www.biomedcentral.com/info/publishing_adv.asp

