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Low risk of recurrence following artesunate—Sulphadoxine— pyrimethamine plus primaquine for uncomplicated *Plasmodium falciparum* and *Plasmodium vivax* infections in the Republic of the Sudan

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Abstract

Background: First-line schizontocidal treatment for uncomplicated malaria in the Republic of the Sudan is artesunate (total dose 12 mg/kg) plus Sulphadoxine/pyrimethamine (25/1.25 mg/kg) (AS/SP). Patients with *Plasmodium vivax* are also treated with 14 days primaquine (total dose 3.5 mg/kg) (PQ). The aim of this study was to assess the efficacy of the national policy.

Methods: Patients above 1 year, with microscopy-confirmed, *Plasmodium falciparum* and/or *P. vivax* malaria were treated with AS/SP. Patients with *P. falciparum* were randomized to no primaquine (Pf-noPQ) or a single 0.25 mg/kg dose of PQ (Pf-PQ1). Patients with *P. vivax* received 14 days unsupervised 3.5 mg/kg PQ (Pv-PQ14) on day 2 or at the end of follow up (Pv-noPQ). Primary endpoint was the risk of recurrent parasitaemia at day 42. G6PD activity was measured by spectrophotometry and the Accessbio Biosensor[™].

Results: 231 patients with *P. falciparum* (74.8%), 77 (24.9%) with *P. vivax* and 1 (0.3%) patient with mixed infection were enrolled. The PCR corrected cumulative risk of recurrent parasitaemia on day 42 was 3.8% (95% CI 1.2–11.2%) in the Pf-noPQ arm compared to 0.9% (95% CI 0.1–6.0%) in the Pf-PQ1 arm; (HR = 0.25 [95% CI 0.03–2.38], p = 0.189). The corresponding risks of recurrence were 13.4% (95% CI 5.2–31.9%) in the Pv-noPQ arm and 5.3% (95% CI 1.3–19.4%) in the Pv-PQ14 arm (HR 0.36 [95% CI 0.1–2.0], p = 0.212). Two (0.9%) patients had G6PD enzyme activity below 10%, 19 (8.9%) patients below 60% of the adjusted male median. Correlation between spectrophotometry and BiosensorTM was low (r_s = 0.330, p < 0.001).

Conclusion: AS/SP remains effective for the treatment of *P. falciparum* and *P. vivax*. The addition of PQ reduced the risk of recurrent *P. falciparum* and *P. vivax* by day 42, although this did not reach statistical significance. The version of the BiosensorTM assessed is not suitable for routine use.

Trial registration https://clinicaltrials.gov/ct2/show/NCT02592408

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Background

There has been a decline in the incidence of malaria in many sub-Saharan countries in the last decade [1–3], which has been attributed to the introduction of artemisinin-based combination therapy (ACT), the scaling up of the distribution of insecticide-impregnated bed nets (ITNs) and indoor residual spraying (IRS) [4]. There has also been an unprecedented increase in the funding of malaria control activities including intensive case management.

In Sudan, the number of malaria cases has declined, although the exact reduction in the burden of disease remains unclear. Information from local studies and reports suggest that in 2002 there were an estimated 9 million clinical cases of malaria and 44,000 deaths [5]. The WHO estimates are lower but highlight a substantial decline between 2000 and 2015, cases falling from 4 million cases in 2000–586,827 cases in 2015, with a total of 868 deaths [4]. However these successes have been associated with an increase in the proportion of infections due to *Plasmodium vivax* [6]. This rise likely reflects malaria control activities mostly targeting *Plasmodium falciparum* [7], and possibly an increased importation of vivax cases from neighbouring countries [6, 8, 9].

Since 2004, the first-line schizontocidal treatment for uncomplicated malaria in the Republic of Sudan has been artesunate plus Sulphadoxine–pyrimethamine (AS/SP) for both *P. falciparum* and *P. vivax* infections [10, 11] (Additional file 1, p. 12–34). In addition primaquine (PQ) is prescribed as a 14 day regimen for the radical cure of *P. vivax* hypnozoites following World Health Organization (WHO) guidelines (total dose 3.5, 0.25 mg/kg daily) [12, 13]; a single dose PQ is not currently recommended for patients with *P. falciparum* as gametocytocidal [14].

Whilst PQ is generally well tolerated, it can cause severe haemolysis in individuals with glucose 6 phosphate dehydrogenase deficiency (G6PDd) [15]. The WHO recommends that where feasible all patients should be tested for G6PD deficiency prior to prescribing primaquine based radical cure, however in view of the lack of available point of care tests to determine G6PDd status, this is not stipulated in the Sudanese anti-malarial treatment policy.

The aim of this study was to assess the efficacy and safety of AS/SP in patients with P. falciparum or P. vivax infection and the additional benefit of PQ either as a single dose for P. falciparum or 14-day regimen for P. vivax. The Biosensor (Accessbio, USA), a novel quantitative point of care G6PD diagnostic was also evaluated.

Methods

Study sites

The study was conducted at two sites in the Republic of Sudan: Gezira Slanj, a semi-urban area north of Khartoum (15°53′11.2″N 32°31′39.9″E), and New Halfa, in the east of the country (15°20′N 35°35′00″E). Malaria transmission in north Khartoum and eastern Sudan is seasonal with the main transmission occurring between September to February [16, 17], although malaria occurs throughout the year at both study sites. Key mosquito vectors in Sudan include *Anopheles arabiensis*, besides *Anopheles gambiae* s.l. and *Anopheles funestus* s.l. [11].

Study population

Study participants were recruited from patients presenting to one of two health care facilities with febrile illness. Patients with peripheral P falciparum and/or P vivax parasitaemia detected by microscopy, aged at least 12 months, with history of fever in the last 24 h or axillary body temperature ≥ 37.5 °C were eligible for enrolment. Patients were excluded if they weighed less than 5 kg, were pregnant, had signs of severe malaria [18], were unable to tolerate oral medication, had a haemoglobin (Hb) concentration less than 8 g/dl, were on regular medication which could interfere with study treatment, or had known allergies to any of the study drugs. Whenever G6PD activity was determined by spectrophotometry participants with G6PD activities below 3.0 U/gHb were excluded from 14 day primaquine treatment [19].

Study design and randomization

The study was designed as an open label, prospective randomized, controlled trial with 42 days of follow up. All patients were treated with AS/SP, allocation to PQ treatment was randomized according to the species of infection. Patients with *P. falciparum* mono-infection were randomized to receive AS/SP either alone (Pf-noPQ) or with a single dose of PQ (Pf-PQ1) administered on day 2. Patients with a *P. vivax* mono or mixed infection were randomized to receive AS/SP either alone (Pv-noPQ) or combined with 14 days of PQ (Pv-PQ14) starting on day 2. Patients in the Pv-noPQ arm received PQ according to national guidelines but delayed until after the end of 42 day follow up.

Treatment allocation was randomized in blocks of 20 and was provided in a sealed opaque envelope, which was only opened by a study nurse once the participant had met all the enrolment criteria and had provided written informed consent. Study staff assigned the patients sequentially according to the sealed envelopes. Blinding of clinical staff towards group allocation was not possible

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with the standard commercially sourced drugs; however, assignment was concealed from the microscopists responsible for determination of the primary endpoint.

Treatment

Quality assured co-formulated blister packs of AS/SP (Shanghai-Sudan Pharmaceutical Ltd, China) were used for the study and patients were treated for 3 days according to national guidelines (target dose 4 mg/kg/day for AS and 25/1.25 mg/kg for SP) [13, 14]. On day 2, patients with *P. falciparum* infection were allocated to receive either no further treatment (Pf-noPQ) or a single dose of 0.25 mg/kg PQ (Pf-PQ1), manufactured by ipca Laboratories Ltd, Mumbai India.

Patients infected with *P. vivax* alone or mixed with *P. falciparum*, were allocated either to AS/SP and 14 days of unsupervised PQ (0.25 mg/kg/day, total dose 3.5 mg/kg) starting on day 2 (Pv-PQ14) or AS/SP alone (Pv-noPQ).

All doses of AS/SP and the single dose PQ in the Pf-PQ1 arm were supervised, however only the first dose of primaquine in the Pv-PQ14 arm was supervised. Study participants were observed for 30 min after administration and those vomiting their dose were treated again with the same dose and observed for a further 30 min.

Patients with recurrent uncomplicated malaria during the follow up period were treated with artemether–lumefantrine (AL) and patients with severe malaria were treated with quinine according to national guidelines [14].

Clinical procedures and follow up

At enrolment demographic data, symptom details and history of anti-malarial medication were collected in a standardized questionnaire. A clinical examination was carried out and up to 5 ml venous blood taken for blood film examination, measurement of haemoglobin concentration (Hb), analysis of G6PD enzyme activity and parasite genotyping. Haemoglobin was measured at the bedside using at Hemocue 201 (Hemocue, Sweden) and G6PD activity measured with an experimental Biosensor™ (Accessbio, USA) following procedures as recommended by the manufacturer and described elsewhere [20, 21].

Patients were examined daily for the first 3 days and thereafter weekly on days 7, 14, 21, 28, 35 and 42. Patients with *P. vivax* were seen on day 16 rather than day 14, at which time a pill count was undertaken. Patients were encouraged to return to the clinic whenever they experienced signs and symptoms consistent with malaria. At each follow up visit, a full physical examination was undertaken, a symptom questionnaire completed, adverse events assessed and capillary blood collected for repeat blood film examination and Hb measurement.

Whenever possible patients who did not return for scheduled follow up visits were contacted and encouraged to return.

Laboratory procedures Microscopy

Thick and thin films were stained with Giemsa and counted per 200 white blood cells (WBC) or 1000 red blood cells (RBC). Presence and density of gametocytes were also assessed per 200WBC. Slides for microscopy were subject to internal and external quality control checks. 20% of all slides were read twice by two local microscopists, and in case of discordant results reread by a third microscopist (internal QC). All slides collected on enrolment and at the time of recurrent parasitaemia were subject to external quality control by a WHO-certified expert microscopist of the Malaria Research Facility of the Papuan Health and Community Development Foundation (PHCDF) in Timika, Papua Province, Indonesia (external QC).

G6PD testing

Due to logistical reasons G6PD activity was only measured at one of the two sites. In New Halfa G6PD activity was measured directly upon sample collection using a handheld Biosensor (AccessBio CareStart, USA) and within 24 h by spectrophotometry (SPINREACT™, Spain) on a Mindray BA-88A (Minddray, China). G6PD normal and deficient controls (Ref: 100 252 0, SPINREACT, Spain), were run daily prior to sample measurement. Patients with *P. vivax* infection and G6PD activity below 3.0 U/gHb (based on spectrophotometry) were excluded from PQ treatment [19].

PCR testing

DNA was extracted from whole blood with a QIAamp DNA blood macro kit (Qiagen) following the manufacturer's recommendations. Extracted DNA was eluted in 50 µl water and used for the genotyping of recurrent *P. falciparum* infections following WHO recommendations [22] by characterizing the length polymorphism of the *msp1*, *msp2*, and *glurp* genes in samples collected at day 0 and on the day recurrent parasitaemia was found.

Recrudescence was determined when, for each marker (*msp1*, *msp2*, and *glurp*), at least one identical-length polymorphism was found between samples collected on day 0 and on the day of recurrent infection. A new infection was defined when, for at least one marker, the length polymorphisms were different between the sample collected on day 0 and that collected on the day of recurrent infection. Samples were defined as indeterminate when no marker could be amplified.

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Study endpoints and definitions

The primary endpoint of the study was the rate of recurrence of peripheral asexual parasitaemia within 42 days of follow up. Secondary endpoints were the proportion of patients with parasitaemia or fever on days 1, 2 and 3 and gametocyte carriage following P. falciparum malaria. Response to treatment was defined according to WHO definitions [23] and treatment failures were categorized as early treatment failures (ETF), late parasitological (LPTF) and late clinical treatment failures (LCTF). Patients without asexual parasitaemia by day 42, and who did not previously meet any of the criteria of ETF, LPTF or LCTF were defined as adequate clinical and parasitological response (ACPR). Fever clearance time (FCT) in patients with a temperature ≥ 37.5 °C at enrolment was defined as the time from drug administration until body temperature (axillary) was below 37.5 °C.

Statistical analysis

Assuming that AS/SP has a 15% risk of treatment failure at day 42 [24], and that this is unaffected by the addition of a single dose of primaquine, a total of 235 patients with P. falciparum infection determine the efficacy of AS/SP within \pm 5% allowing for a loss to follow up rate of 10%. To achieve 80% power, 95% significance level to detect a decrease in P. vivax recurrence from 30 to 5%, following AS/SP plus or minus 14 days of primaquine (26), a sample size of 40 patients would be needed in each treatment arm.

Data were entered using EpiData (version 3.1,EpiData Software, Denmark) and analysed using Stata version 14 (Stata Corp., USA). The primary analysis was an intention to treat (ITT) analyses including all participants enrolled into the study. A secondary analysis was done using a modified intention to treat (mITT) approach in which patients who had received sub-optimal treatment dosage were excluded. Sub-optimal drug dosage was defined as a total dose below 4 mg/kg for AS, below 25 mg/kg for Sulphadoxine and 1.25 mg/kg for pyrimethamine [13, 14]. The sub-optimal PQ dose was defined as below 0.15 mg/kg in the Pf-PQ1 arm and below 2.5 mg/kg total dose of primaquine in the Pv-PQ14 arm [25].

Normally distributed data were compared using Student's t-test, the t-test for paired samples or one-way analysis of variance, and non-parametric comparisons were made using the Mann–Whitney U test or Wilcoxon signed rank test for paired samples. Proportions were examined using χ^2 with Yates' correction or Fisher's exact test. Correlations were assessed using the Pearson test for correlated proportions for normal distributed variables and the Spearman rank test for non-normal distributed variables. Efficacy endpoints were assessed by survival analysis, in which the cumulative risk of failure was

calculated by the Kaplan–Meier product limit formula [26]. The equality of survivor functions was tested using the log-rank test (exponential scores test).

The results of the survival analysis were adjusted according to parasite genotyping results of the admission and recurrent parasitaemia. Patients with recurrent parasitaemia due to a different species identified on admission or a PCR confirmed re-infection were censored at the day of occurrence. Following a conservative approach indeterminate PCR results were considered treatment failures. Parasite clearance was presented as the proportion of patients with microscopy negative results within the first 3 days, missing data were treated as suggested by the World Wide Antimalarial Resistance Network (WWARN) [27].

Safety analysis included the risks of a drop in Hb concentration greater than 25%, severe anaemia (Hb < 7 g/dl), requiring a blood transfusion, and the proportion of patients with adverse and serious adverse events. Haematological response was assessed from the fractional fall in Hb between baseline and days 2, 7 and 16. The effect of malaria species, peripheral parasitaemia, G6PD activity and total dose of primaquine on the degree of haemolysis was assessed by multivariable linear regression using a backward selection of variables [28]. The performance of the Biosensor was assessed using standard formulae [29, 30] in which the adjusted male median (AMM) was considered 100% G6PD activity [15].

Results

Study profile and baseline characteristics

Between 7th December 2015 and 9th March 2016, 4887 patients were screened for malaria. Overall 486 (9.9%) patients had malaria, of whom 309 (63.6%) were enrolled in the study (213 [69.9%] in New Halfa and 96 [31.1%] in Gezira Slanj) (Fig. 1). The main reason for non-enrolment of patients with malaria, was declining to participate in the study (113/177, 63.8%). Other reasons are listed in Fig. 1. No patient was excluded from 14 day primaquine treatment because of low G6PD activity as measured by spectrophotometry.

Based on microscopy a total of 231 (74.8%) patients were infected with *P. falciparum*, 77 (24.9%) with *P. vivax* and one patient (0.3%) with a mixed infection of *P. falciparum* and *P. vivax*. Patients with *P. falciparum* infections had a significantly higher parasitaemia and were less likely to be gametocytaemic at enrolment than those with *P. vivax* infection (Table 1). Baseline characteristics between the randomized groups for each species were similar, proportions of patient enrolled into each arm were similar per site (Table 1).

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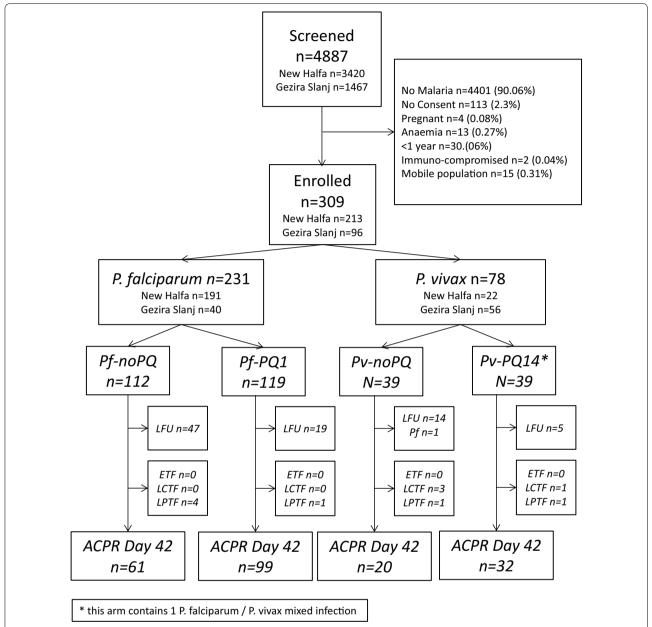


Fig. 1 Study profile. *ETF* early treatment failure, *LCTF* late clinical treatment failure, *LPTF* late parasitological treatment failure, *ACPR* adequate clinical and parasitological response

The target dose of AS was achieved in 100.0% patients, but only 59.5% (184/309) of patients received the target dose of S/P (Table 2). The median dose of PQ prescribed for patients in the Pf-PQ1 arm was 0.25 mg/kg (IQR 0.192–0.333) with 5 patients (4.2%) receiving less than 0.15 mg/kg. The median total dose of PQ prescribed in the Pv-PQ14 arm was 4.20 mg/kg (IQR 3.18–4.67), with 15.4% (6/39) patients prescribed a total dose less than 2.5 mg/kg [26]. Pill counts were undertaken in all patients

receiving 14 days unsupervised PQ (Pv-PQ14) at the end of their treatment course. One participant (2.6%) had only taken 2 of the 14 doses of PQ, but no pills were remaining for any of the other patients.

Early therapeutic response

Of the 231 patients with *P. falciparum* infection, 74.7% (130/174) were still parasitaemic on day 1, and this fell to 4.6% (8/174) on day 2. One 10-year old patient with

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Table 1 Baseline characteristics

	P. falciparum				P. vivax ^a				
	AII N=231	Pf-noPQ N=112	Pf-PQ1 N=119	р	AII N=78	Pv-PQ14 N=39	Pv-noPQ N=39	р	
N (%) male	117 (50.6%)	53 (47.3%)	65 (53.7%)	0.329	40 (51.3%)	20 (51.3%)	20 (51.3%)	1.000	
New Halfa (%)	191 (100.0)	92 (48.2)	99 (51.8)	0.833	22 (100.0)	12 (54.5)	10 (45.5)	0.615	
Gezira Slanj (%)	40 (100.0)	20 (50.0)	20 (50.0)		56 (100.0)	27 (48.2)	29 (51.8)		
Median age in years, (IQR)	19.0 (12.0–28.0)	18.5 (12.0–28.0)	19.0 (12.0–28.0)	0.310	17.0 (10.0–27.0)	16.0 (10.0–23.0)	19.0 (9.0–30.0)	0.272	
0–5 years, N (%)	18 (7.8%)	8 (44.4%)	10 (55.5%)	0.824	4 (5.1%)	3 (75.0%)	1 (25%)	0.615	
> 5 years, N (%)	213 (92.2%)	104 (48.8%)	109 (51.2%)		74 (94.1%)	36 (48.6%)	38 (51.4%)		
Body Temp. mean °C (95% CI)	38.8 (38.7–38.8)	38.8 (38.7–38.9)	38.8 (38.8–38.9)	0.499	38.6 (38.4–38.7)	38.5 (38.4–38.7)	38.5 (38.4–38.7)	0.415	
Median Hb in g/ dl (IQR)	11.7 (10.8–12.8)	11.7 (10.7–12.8)	11.8 (10.9–12.8)	0.535	11.0 (10.4–13.0)	12.0 (10.9–13.0)	11.0 (10.0–12.3)	0.113	
Anaemia (< 10 g/ dl), N (%)	13 (5.6%)	8 (7.1%)	5 (4.2%)	0.332	8 (10.3%)	4 (10.3%)	4 (10.3%)	1.000	
Weight, mean kg (95% CI)	57.4 (54.0–60.7)	56.7 (51.8–61.6)	58.0 (53.2–62.7)	0.357	47.2 (42.4–52.1)	44.3 (38.3–50.3)	50.1 (42.3–58.0)	0.119	
Parasitaemia (µl ⁻¹), geomet- ric mean (95% CI)	7840 (7443–8259)	7779 (7226–8375)	7898 (7333–8508)	0.321	6939 (6383–7544)	6933 (6164–7797)	6947 6135–7865)	0.956	
Gametocytaemia (µl ⁻¹), geomet- ric mean (95% CI)	0	0	0	1.000	252 (224–284)	245 (203–296)	259 (222–303)	0.449	
Gametocytaemia, N (%)s	0 (0.0%)	0 (0.0%)	0 (0.0%)	1.000	78 (100.0%)	39 (100.0%)	39 (100.0%)	1.000	
Mean G6PD activity (U/gHb) (95% CI)	9.95 (9.52–10.38)	10.06 (9.37–10.74)	9.85 (9.30–10.39)	0.316	9.57 (8.27–10.88)	9.79 (7.85–11.73)	9.40 (7.35–11.44)	0.381	

^a Includes one patient with *P. falciparum/P. vivax* mixed infection

Table 2 Treatment provided (in mg/kg bodyweight)

	Artesunate	Sulphadoxine	Pyrimethamine	Primaquine ^b
Target dose [13]	4 mg/kg per day for 3 days (range 2–10 mg/kg)	25 mg/kg single dose (range 25–70 mg/kg)	1.25 mg/kg single dose (range 1.25–3.5 mg/kg)	0.25 mg/kg or 3.5 total dose
P. falciparum (n = 231)				
Mean total dose received in mg/kg (95% CI)	10.3 (9.77–10.74)	34.26 (31.37–37.15)	1.7 (1.57–1.86)	0.28 (0.254–0.305)
Median total dose received in mg/kg (IQR, range)	9. 23 (7.69–12.00, 5.36–25.00)	25.42 (19.23–41.67, 13.39–115.38)	1.3 (0.96–2.08, 0.67–5.77)	0.25 (0.192–0.333, 0.000–0.789)
N (%) patients with suboptimal dosage ^a	0 (0.0%)	104 (45.0%)	104 (45.0%)	5 (4.2)
P. vivax/mixed infection (n = 78)				
Mean total dose received in mg/kg (95% CI)	10.9 (10.32–11.64)	39.3 (34.76–43.89)	2.0 (1.74–2.2.)	4.2 (3.57–4.86)
Median total dose received in mg/kg (IQR, range)	10.81 (9.23–7.06, 6.12–27.78)	30.0 (23.07– 55.56, 15.31– 111.11)	1.5 (1/15–2.78, 0.77–5.56)	4.2 (3.18–4.67, 0.12–11.67)
N (%) patients with suboptimal dosage ^a	0 (0.0%)	21 (26.9)	21 (26.9)	6 (15.4%)

^a Suboptimal dosage was defined as a total dose below 4 mg/kg AS, below 25 mg/kg sulphadoxine, below 1.25 mg/kg pyrimethamine and below 0.15 mg/kg (Pf) or 2.5 mg/kg (Pv) primaquine

 $[^]b \ \ \text{Only patients from Pf-PQ arm (n=119, 0.25 mg/kgBW) and Pv-PQ14 (n=39, 3.5 mg/kgBW) arm included a substitution of the property of t$

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a baseline parasitaemia of 2750 parasites/ μ l was afebrile, but still parasitaemic (475 μ l⁻¹) on day 3; he was subsequently lost to follow up. Fever clearance in all other patients with *P. falciparum* infection was rapid, with 95.4% (124/130) patients being afebrile by day 2.

Parasite clearance in patients with *P. vivax* was slower than that for *P. falciparum*. Of the 78 patients with *P. vivax* or mixed infection, 96.8% (60/62) were still parasitaemic on day 1 and 17.7% (11/62) on day 2. One 35-year old patient (1.6%) with a baseline parasitaemia of 9300 parasites/ μ l was afebrile, but remained parasitaemic (645 μ l⁻¹) on day 3; the patient was subsequently lost to follow up. All patients with *P. vivax* parasitaemia had gametocytes present at baseline, but by day 2 this proportion had fallen to 1.6% (1/62). Fever clearance was fast with only one patient with *P. vivax* being febrile by day 2.

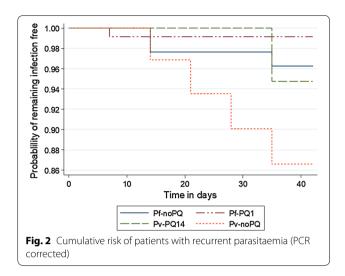
Efficacy outcomes

A total of 85 (27.5%) patients were lost to follow up before the end of the study and one patient (0.32%) presented with *P. falciparum* infection on day 28 after being enrolled with a *P. vivax* infection (Fig. 1). There were no early treatment failures in any treatment arm, but 11 (3.6%) patients represented with late treatment failure (LTF), 7 of which were classified as LPTF and 4 as LCTF (Fig. 1).

Plasmodium falciparum arms

In the *P. falciparum* arm, a total of 5 patients had recurrent parasitaemia on days 7 (n=2), 14 (n=2) and 35 (n=1). The overall risk of recurrent parasitemia on day 28 was 2.0% [95% CI 0.75–5.19%], the risk being greater in patients treated with Pf-noPQ (3.4% [95% CI 1.1–10.3%]) compared to 0.9% (95% CI 0.1–6.0%) in those treated with Pf-PQ1; Hazard Ratio HR=0.26 [95% CI 0.03–2.47], p=0.203.

By day 42 the risk of parasite recurrence was 2.5% [95% CI 2.06-6.01%]; 4.8% (95% CI 1.8-12.4%) in the Pf-noPQ and 0.9% (95% CI 0.1-6.0%) in the Pf-PQ14 (HR=0.19 [95 CI 0.02-1.67], p=0.092); Fig. 2. PCR demonstrated one patient with a reinfection in the Pf-noPQ arm, with the 4 other recurrent samples of the Pf-PQ1 arm providing indeterminate results. The PCR adjusted risk of failure on day 28 was 2.4% (95% CI 0.6-0.9%) and 3.8% (1.2-11.2%) on day 42 (HR=0.25 [95 CI 0.03-2.38], p=0.188) for the Pf-noPQ arm. Site specific efficacy estimates are summarized in Table 3, efficacy between sites for the individual arms did not vary significantly (all p>0.05). No patients initially infected with *P. falciparum* had gametocytes detected on blood film examination during follow up.



Plasmodium vivax arms

In the *P. vivax* arms, a total of 6 patients had recurrent parasitaemia at days 14 (n=1), 21 (n=1), 28 (n=1), 35 (n=3). In patients not receiving 14 days primaquine the risk of recurrent *P. vivax* at day 28 was 9.9% [95% CI 3.3–27.8%] compared to 0% in those treated with AS/SP plus 14 days primaquine; p=0.046. At day 42 the respective risks had risen to 13.4% [95% CI 5.2–31.9%] in the Pv-noPQ arm and 5.3% [95% CI 1.3–19.4%] in the Pv-PQ14 arm; HR 0.36, [95% CI 0.1–2.0], p=0.212 (Fig. 2). There was no difference in treatment efficacy between sites (Table 3).

Modified intention to treat analysis and optimal versus suboptimal treatment

In the mITT analysis, 41.7% (129/309) of patients receiving a sub-optimal dose of any of the study drugs were excluded, leaving 127 (54.9%) of patients with *P. falciparum* and 53 (67.9%) patients with *P. vivax* (Table 3). The overall risk of recurrent parasitaemia was lower in patients who had received optimal treatment, although this did not reach statistical significance in any arm by day 42.

No patient received sub-optimal AS (Table 2). For patients of the Pv-noPQ arm who received less than 25 mg/kg SP, the risk of recurrence was 39.4% [95% CI 16.90–74.20] compared to 0.0% in patients of the same arm who received higher treatment doses (p=0.003). No significant differences in efficacy were found when comparing sub-optimal versus optimal SP treatment in the other treatment arms.

Haematological response

At baseline the median Hb concentration was 11.7 g/dl in patients with *P. falciparum* and 11.0 g/dl in patients

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Table 3 Failure rates overall and per site on day 28 and 42 using the ITT and mITT populations

	P. falciparum (PCR corrected)				P. vivax			
	Pf-noPQ	Pf-PQ1	p value	Hazards ratio	Pv-noPQ	Pv-PQ14	p value	Hazards ratio
ITT								
D28								
Overall (95% CI)	2.35 (0.59–9.08)	0.86 (0.12–5.96)	0.419	0.385 (0.035– 4.241)	9.93 (3.31–27.76)	0.00	0.046	-
New Halfa (95% CI)	2.90 (0.73–11.10)	1.03 (0.15–7.09)	0.406	0.374 (0.034– 4.129)	20.0 (3.08–79.62)	0.00	0.157	-
Gezira Slanj (95% Cl)	0.00	0.00	1.000	-	8.17 (2.11–28.92)	0.00	0.120	-
p value ^b D42	0.493	0.658	_	-	0.487	1.000	-	
Overall (95% CI)	3.75 (1.22–11.22)	0.86 (0.12–5.96)	0.189	0.247 (0.026– 2.378)	13.39 (5.24– 31.91)	5.26 (1.34–19.44)	0.212	0.358 (0.066– 1.956)
New Halfa (95% Cl	4.66 (1.52–13.85)	1.03 (0.15–7.09)	0.176	0.238 (0.025– 2.293)	20.0 (3.08–79.62)	0.00	0.157	_
Gezira Slanj (95% Cl)	0.00	0.00	1.000	-	12.35 (4.15– 33.59)	7.14 (1.84–25.65)	0.480	0.535 (0.089– 3.200)
p value ^b	0.389	0.658	-	_	0.392	0.684	-	_
mITT ^a								
D28								
Overall (95% CI)	2.00 (0.28–13.36)	0.00	0.282	_	0.00	0.00	1.000	_
New Halfa (95% CI)	2.86 (0.41–18.60)	0.00	0.237	_	0.00	0.00	1.000	_
Gezira Slanj (95% Cl)	0.00	0.00	1.000	_	0.00	0.00	1.000	-
p value ^b	0.513	1.000	-	=	1.000	1.000	-	=
D42								
Overall (95% CI)	4.28 (1.08–16.10)	0.00	0.117	-	0.00	6.90 (1.77–24.86)	0.247	-
New Halfa (95% CI)	6.21 (1.58–22.73)	0.00	0.083	-	0.00	0.00	1.000	-
Gezira Slanj (95% Cl)	0.00	0.00	1.000	-	0.00	10.00 (2.60– 34.40)	0.186	_
p value ^b	0.340	1.000	-	=	1.00	0.334	0.334	=

Significant values are in italics (p < 0.05)

with *P. vivax* (Table 1). The greatest median fractional fall in Hb occurred in the *P. falciparum* arms between baseline and day 2 prior to administration of any primaquine, whereas there was no median drop in Hb among patients with a *P. vivax* infection (Table 4). On day 2 patients infected with *P. falciparum* had a median fractional change in Hb of -4.9% (IQR -10.6 to +0.9, range -29.3 to +50.7), compared to patients infected with *P. vivax* whose Hb did not change (IQR -7.1 to +7.7 to 4.6, range -23.1 to +50.0) over the same period; p=0.015 (Fig. 3). A total of 1.7% (4/231) of patients with *P. falciparum* had a fractional fall in Hb greater than 25%

between day of enrolment and day 2, while this was not the case for any patient with P. vivax infection (p = 0.316).

The median Hb levels increased in all treatment arms between days 0 and 7 (Fig. 3, Table 4). In patients with *P. falciparum* the Hb concentration at day 7 had risen 6.0% (IQR — 0.8 to 14.8) from baseline following treatment with Pf-noPQ compared to 13.2% (IQR 0.0 to 23.9) after treatment with Pf-PQ1; p=0.007. In patients with *P. vivax*, there was no change in Hb (IQR 0.0 to 10.0) between day 0 and day 7 following treatment with Pv-noPQ compared to 9.1% (IQR 0.0 to 15.4) in vivax patients treated with primaquine (Pv-PQ14);

^a mITT population excluded 132 (42.7%) with incomplete or suboptimal dosing of study drugs

^b This p value refers to site specific differences and does not consider overall results

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Table 4 Relative change in the mean Hb concentration between baseline and day of follow up

Relative median change in Hb from baseline	All	P. falciparum			All	P. vivax		
		Pf-noPQ	Pf-PQ1	р		Pv-noPQ	Pv-PQ14	р
D1 (IQR, n)	- 2.7% (- 9.09 to 1.92, 174)	- 3.8% (- 7.75 to 2.27, 86)	- 2.42% (- 9.33 to 0.00, 88)	0.8161	0.0% (- 5.30 to 0.92, 62)	0.0% (- 4.72 to 0.00, 30)	0.0% (- 5.68 to 8.71,	0.219
D2 (IQR, n)	- 4.9% (- 10.62 to - 0.87, 163)	- 4.4% (- 9.05 to 0.43, 72)	- 5.6% (- 11.9 to 2.65, 91)	0.525	0.0% (- 7.14 to 7.69, 61)	0.0% (- 7.14 to 1.63, 32)	0.0% (- 7.69 to 9.09, 29)	0.460
D7 (IQR, n)	11.2% (0.00 to 19.54, 164)	6.0% (- 0.76 to 14.81, 62)	13.2% (0.00 to 23.89, 102)	0.007	8.0% (0.00 to 15.38, 56)	0.0% (0.00 to 10.00, 25)	9.1% (0.00 to 15.38, 31)	0.106
D14 (IQR, n)	16.5% (4.27 to 27.08, 162)	13.3% (0.00 to 22.69, 57)	18.6% (7.69 to 30.08, 105)	0.006	NA	9.1% (NA, 1)	n=0	NA
D16 (IQR, n)	n=0	n=0	n=0	NA	7.7% (0.00 to 18.18, 67)	7.7% (0.00 to 18.18, 30)	7.7% (0.00 to 18.18, 37)	1.000
D21 (IQR, n)	15.4% (0.85 to 23.08, 147)	15.0% (0.00 to 26.85, 61)	15.4% (4.07 to 22.76, 86)	0.939	8.3% (0.00 to 18.18, 55)	7.7% (0.00 to 18.18, 23)	8.7% (0.00 to 18.18, 32)	0.614
D28 (IQR, n)	16.9% (4.41 to 26.46, 140)	13.5% (0.83 to 24.77, 62)	17.7% (6.40 to 28.46, 78)	0.284	11.1% (0.00 to 21.10, 47)	10.0% (0.00 to 18.18, 21)	12.5% (0.00 to 21.10, 26)	0.754

Significant values are in italics (p < 0.05)

p = 0.106. None of the patients included required a blood transfusion.

Tolerability

A total of 551 adverse events (AE) were reported, of which 497 (90.2%) were categorized as mild and 51 (9.8%) as moderate. Three (0.5%) mild events were recorded as possibly related to AS/SP. The most frequent AEs were gastrointestinal symptoms including poor appetite (40.3%), nausea (25.6%) and abdominal pain (20.7%) (Table 5). In 93.8% (226/241) of cases the first episode of gastrointestinal symptoms was reported on or after day 2. In total 68.9% (82/119) of patients treated with Pf-PQ1 reported at least one gastrointestinal AE on or after day 2 compared to 81.3% (91/112) of those with Pf-noPQ; relative risk = 0.820 [95% CI 0.631-1.065], p = 0.155. The corresponding relative risk in patients from the Pv-PQ14 arm compared to patients from the Pv-noPQ arm was 0.943 [95% CI 0.592-1.50], p=0.808. No serious adverse events occurred in any study patient.

G6PD activity and performance of the biosensor (AccessBio–CareStart)

G6PD activity was quantified by spectrophotometry and BiosensorTM in 68.5% (213/309) patients. The adjusted male median by spectrophotometry was 9.7 U/gHb, compared to 7.9 U/gHb derived by the BiosensorTM (p<0.001). By spectrophotometry 2 (0.9%) patients with

P. falciparum infection had G6PD activities less than 10% of the AMM, no patient had G6PD activities between 10 and 30% and 18 (8.5%) patients had G6PD activity between 30 and 60% (Fig. 4).

The Biosensor[™] identified 1 patient with less than 10% G6PD activity, 11 participants (5.2%) with activities between 10 and 30% activity, and 27 patients with G6PD activity between 30 and 60% (Table 6, Fig. 5). None of the patients with G6PD activities below 10% were identified by the biosensor correctly. The correlation between the activity recorded by the two assays was r_s = 0.330 (p < 0.001), with a mean difference of 2.1 U/gHb and the 95% limit of agreement (LoA) ranged from − 5.1 U/gHb to 9.2 U/gHb (Figs. 4, 5). In the absence of true G6PD deficient results with activities < 30% of the AMM sensitivity could not be calculated, while specificity was 0.94 (95% CI 0.90−0.97). Sensitivity and specificity at 60% of the AMM were 0.55 (95% CI 0.32−0.77) and 0.86 (95% CI 0.80−0.91), respectively.

Discussion

This efficacy study demonstrates that when the target dose of AS/SP was achieved the risk of recurrent parasitaemia was low for patients with either *P. falciparum* or *P. vivax* in the two Sudanese study sites assessed. The dosage scheme applied followed recommendations of the WHO; never the less almost half of the patients received a dose of SP less than the intended target dose of 25 mg/

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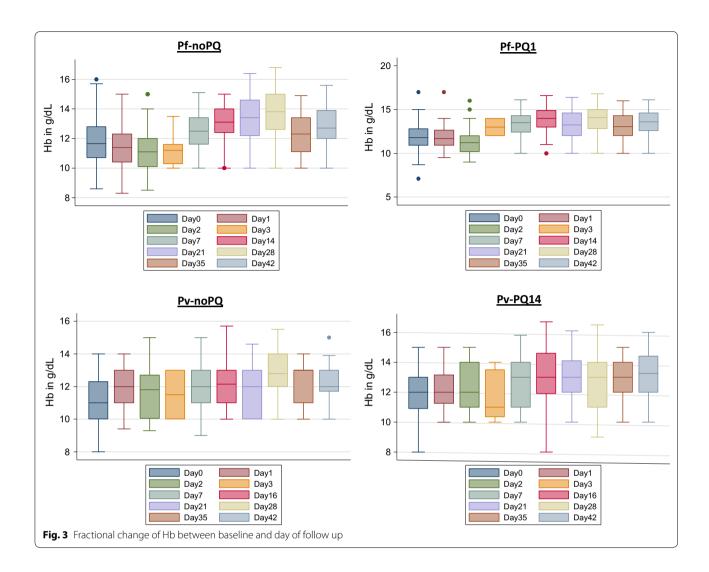


Table 5 Overview of main adverse events

Type of AE	Total	P. falciparum			P. vivax		
		Pf-noPQ	Pf-PQ1	р	Pv-noPQ	Pv-PQ14	р
Poor appetite, n (%)	222 (40.3)	93 (47.2)	75 (35.7)	0.012	30 (41.1)	24 (33.8)	0.366
Nausea, n (%)	145 (25.6)	58 (29.4)	60 (28.6)	0.533	13 (17.8)	14 (19.7)	0.769
Abdominal pain, n (%)	114 (20.7)	29 (14.7)	51 (24.3)	< 0.001	21 (28.8)	13 (18.3)	0.140
Vomiting, n (%)	37 (6.7)	15 (7.6)	13 (6.2)	0.536	5 (6.8)	4 (5.6)	0.518
Rash, n (%)	1 (0.2)	0 (0.0)	1 (0.5)	0.521	0 (0.0)	0 (0.0)	1.000
Other, n (%)	36 (6.5)	2 (1.0)	14 (6.7)	0.004	4 (5.5)	16 (22.5)	0.004
Total, n (%)	551	197	210		73	71	

Significant values are in italics (p < 0.05)

kg [13]. While the target dose is designed for patients with a maximum body weight of 60 kg, a substantial fraction of the study population weighed more, resulting in under dosing (Additional file 1) and in patients with *P. vivax* treated with AS/SP alone, this was associated

with an almost 40% risk of recurrence by day 42. The additional administration of single dose primaquine in patients with *P. falciparum* and 14 days primaquine in *P. vivax*, was associated with lower risks of parasite recurrence (5.3- and 2.8-fold, respectively), although these

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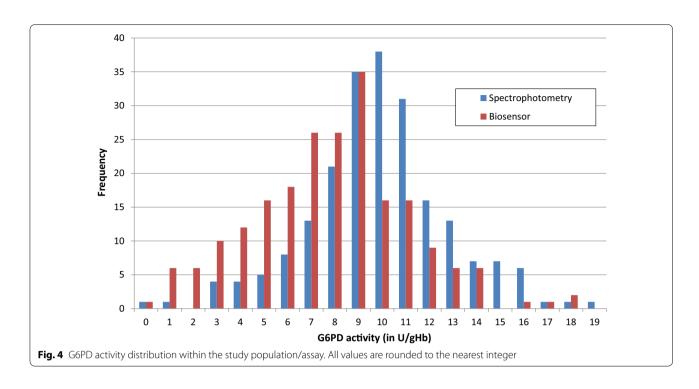


Table 6 Categorized G6PD result/assay

G6PD cut off	Biosensor [™]						
	<10%	10 to <30%	30 to <60%	≥ 60%			
Spectrophotom	etry						
< 10 %	0	0	2	0	2		
10 to < 30 %	0	0	0	0	0		
30 to < 60 %	0	2	7	9	18		
≥60%	1	9	17	166	193		
Total	1	11	27	176	213		

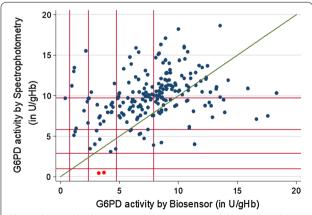


Fig. 5 Scatter plot biosensor result versus spectrophotometry. Green line = line of equality, red lines from origin outwards represent 10, 30, 60 and 100% G6PD activity/assay, red markers = subjects with < 30% activity, not recognized by the Biosensor

benefits did not reach statistical significance. Results on gametocyte carriage are limited by the small denominator applied for microscopy (200WBC).

In 2006, a study in the east of the country reported a day 28 PCR corrected efficacy of 93.5% for AS/SP for *P. falciparum* [31]. Other clinical studies conducted between 2010 and 2015 at six sites suggest that the PCR corrected efficacy of AS/SP against *P. falciparum* by day 28 generally exceeds 95%, except for one site in Gedaref, in the East of the country, where the efficacy fell from 91% in 2012 to 87% in 2015 [32].

One study from Kassala, Eastern Sudan reported a PCR corrected efficacy of 93.7% by day 28 [33], while a study from Damazin, South-Eastern Sudan reported a PCR corrected efficacy of 80% for AS/SP by day 28 [34]. The study in Kassala also identified a high proportion of parasites with double mutations in the *pfdhfr* and *pfdhps* genes and a single case with a triple mutation in the *pfd-hfr* gene, all associated with antifolate resistance [33].

In Somalia, the efficacy of AS/SP has fallen to 88% and artemether–lumefantrine has replaced AS/SP as the first-line treatment for *P. falciparum* [35]. In March 2017, after patient enrolment to the current study had been completed, North Sudan anti-malarial policy was also changed from AS/SP to artemether–lumefantrine (AL) for the treatment of uncomplicated malaria.

In areas co-endemic for *P. falciparum* and *P. vivax* AS/SP has been used as a universal schizontocidal treatment for both species particularly in locations where

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diagnostic facilities are unable to distinguish reliably between species. To date 3 published trials have assessed the efficacy of AS/SP against P. vivax. Titra et al. found AS/SP to have almost 90% efficacy against P. vivax in Papua, Indonesia [36]. However clinical trials with longer follow up revealed that the risk of recurrent P. vivax by day 42 after AS/SP rose to 25% in Afghanistan [37], and 67% in Papua New Guinea [38]. In the current study, the overall risk of P. vivax recurrence at day 42 was 13.4% in patients treated with AS/SP alone, less than half that expected in our a priori power calculation. The risk of recurrence fell to 5.3% in those treated with AS/SP plus 14 days primaquine although this did not reach statistical significance (p = 0.212); Table 3. A longer follow-up and larger sample size would be required to document the additional benefits of primaquine radical cure that appeared to be emerging [39].

WHO guidelines recommend that the radical cure of *P. vivax* can be achieved with a 14-day regimen of primaquine, although the long course of treatment is associated with poor adherence and compromised effectiveness [25, 40–44]. In some settings unsupervised PQ appears to work well [45], and indeed in the current study pill counting, undertaken on day 16, suggested excellent adherence to the 14-day treatment regimen, with only one patient out of 39 failing to take all of their tablets. However, the nature of a clinical trial including regular review may influence patients behaviour significantly, leading to an overestimation of the true clinical effectiveness in non-trial settings.

PQ is known to cause haemolysis in G6PD deficient individuals with the nadir usually observed 2-7 days after initial exposure [46]. The greatest fall in Hb was observed at day 2 prior to the administration of any primaguine and thus is likely to be attributable to parasite rather than drug induced haemolysis. In patients with P. falciparum infection there was no further fall in Hb observed between day 2 and day 7, with or without a single dose of primaguine. These observations are reassuring and concur with previous reports from Tanzania [47] supporting the current WHO guidelines that do not recommend routine G6PD testing prior to the administration of a single dose of PQ [13]. None of the patients with P. vivax infection had G6PD activity below 30%, although 2/22 (9.1%) had intermediate activity between 30 and 60%, reassuringly none of the patients had a significant fall in Hb after day 2.

Howes et al. had earlier documented a country wide G6PD deficiency prevalence of 15.3% with higher frequencies towards the West of the country and lower frequencies in the East [48]. These prevalence estimates are based on 8 surveys and a definition of G6PD deficiency using a cut-off of 30% [15]. G6PD enzyme activity was

only assessed in patients enrolled in New Halfa in the East of the country and defined following the same cutoff. Based on this definition only two patients (0.9%) were
G6PD deficient. This lower prevalence is partly explained
by the location of sample collection which was close to
the borders with Eritrea and Ethiopia, where the prevalence of G6PD deficiency is low [48]. Furthermore, it is
likely that G6PD deficiency offers some degree of protection against clinical malaria [49–54], and the patients
with deficiency identified in cross sectional surveys may
be less likely to present to clinics with symptomatic
illness

The study also evaluated a novel point of care BiosenorTM (Accessbio/Carestart, USA) which was compared to the reference method of spectrophotometry. The results highlight that the current BiosensorTM underdiagnosed severe G6PD deficiency, a finding consistent with a recent similar study conducted in Bangladesh [21]. Out of the 21 participants with G6PD activities below 60% by spectrophotometry only 11 (52.4%) were correctly classified with both patients with severe G6PDd classified as intermediate deficiency using the BiosensorTM.

Conclusion

AS/SP appears to retain adequate efficacy in uncomplicated malaria in New Halfa and Gezira Slanj. Both a single dose of primaquine in patients with P. falciparum and a 14-day regimen in patients with P. vivax were well tolerated, and associated with a modest reduction in the number of recurrences by day 42, however reduction was not significant for P. falciparum. The Biosensor achieved suitable operational characteristics, but test performance is insufficient for routine diagnosis. A second generation with integrated haemoglobin measurement will be introduced to the market and may provide a more suitable solution.

Additional files

Additional file 1. Scatter plot body weight vs. total dose SP received. red line = target dose recommended by the WHO.

Additional file 2. Database.

Authors' contributions

MMAH, KT, RNP and BL devised the experiments; BL wrote the first draft; BL, KT RG and RNP analysed the data; MMAH and NSM supervised all laboratory analysis; JM performed parasite genotyping; all authors revised the manuscript and provided scientific input in writing the manuscript. All authors read and approved the final manuscript.

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Competing interests

The authors declare that they have no competing interests.

Availability of data and materials

All data published are included in Additional file 2 linked to this article.

Ethics approval and consent to participate

The study was approved by the Human Research Ethics Committee of the Northern Territory Department of Health and Menzies School of Health Research, Australia (HREC:2015-2493), the University of Khartoum Institute of Endemic Diseases Research Ethics Committee (Ref. No.: 02/10/2015) and State Ministry of Health, Sudan. The study was registered at clinicaltrials.gov (NCT02592408). Prior to enrolment written informed consent was collected from all participants, or in the case of those under 18 years old from their legal guardian. Written assent was collected from all patients aged 11–18 years old.

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References

- Bhattarai A, Ali AS, Kachur SP, Martensson A, Abbas AK, Khatib R, et al. Impact of artemisinin-based combination therapy and insecticidetreated nets on malaria burden in Zanzibar. PLoS Med. 2007;4:e309.
- Ceesay SJ, Casals-Pascual C, Erskine J, Anya SE, Duah NO, Fulford AJ, et al. Changes in malaria indices between 1999 and 2007 in The Gambia: a retrospective analysis. Lancet. 2008;372:1545–54.
- O'Meara WP, Bejon P, Mwangi TW, Okiro EA, Peshu N, Snow RW, et al. Effect of a fall in malaria transmission on morbidity and mortality in Kilifi, Kenya. Lancet. 2008;372:1555–62.
- WHO. World Malaria Report 2016. Geneva, World Health Organization, 2016.
- Abdalla SI, Malik EM, Ali KM. The burden of malaria in Sudan: incidence, mortality and disability—adjusted life—years. Malar J. 2007;6:97.
- Mahgoub H, Gasim GI, Musa IR, Adam I. Severe Plasmodium vivax malaria among sudanese children at New Halfa Hospital, Eastern Sudan. Parasit Vectors. 2012;5:154.
- NMCP. Sudan malaria programme performance review—aide memoire. Khartoum: NMCP; 2013.
- Kebede S, Medhin G, Berhe N, Teklehaymanot T, Gebru T, Clause R, et al. Return of chloroquine-sensitive *Plasmodium falciparum* parasites and emergence of chloroquine-resistant *Plasmodium vivax* in Ethiopia. Malar J. 2014:13:244

- Ketema T, Bacha K. Plasmodium vivax associated severe malaria complications among children in some malaria endemic areas of Ethiopia. BMC Public Health. 2013;13:637.
- Abdelgader TM, Ibrahim AM, Elmardi KA, Githinji S, Zurovac D, Snow RW, et al. Progress towards implementation of ACT malaria case-management in public health facilities in the Republic of Sudan: a cluster-sample survey. BMC Public Health. 2012;12:11.
- 11. Republic of Sudan, National Ministry of Health. Five years Strategic Plan for the National Malaria Control Programme. Khartoum: NMCP; 2010.
- Graves PM, Gelband H, Garner P. Primaquine or other 8-aminoquinoline for reducing *Plasmodium falciparum* transmission. Cochrane Database Syst Rev. 2015;2:CD008152.
- WHO. Guidelines for the treatment of malaria. Geneva: World Health Organization; 2015.
- Ibrahim HMO, Ali FA, Elmardi KA. The national protocol for treatment of malaria. Khartoum: Republic of Sudan, National Ministry of Health; 2010.
- von Seidlein L, Auburn S, Espino F, Shanks D, Cheng Q, McCarthy J, et al. Review of key knowledge gaps in glucose-6-phosphate dehydrogenase deficiency detection with regard to the safe clinical deployment of 8-aminoquinoline treatment regimens: a workshop report. Malar J. 2013;12:112.
- Himeidan YE, Elbashir MI, el El-Rayah A, Adam I. Epidemiology of malaria in New Halfa, an irrigated area in eastern Sudan. Eastern Mediterr Health J. 2005;11:499–504.
- El Sayed BB, Arnot DE, Mukhtar MM, Baraka OZ, Dafalla AA, Elnaiem DE, et al. A study of the urban malaria transmission problem in Khartoum. Acta Trop. 2000;75:163–71.
- Severe falciparum malaria. World Health Organization, communicable diseases cluster. Trans R Soc Trop Med Hyg. 2000;94(Suppl 1):S1–90.
- Spinreact. Glucose-6-phosphate dehydrogenase, enzymatic-UV. Spain: Spinreact; 2014. p. 1.
- Ley B, Alam MS, O'Donnell JJ, Hossain MS, Kibria MG, Jahan N, et al. A comparison of three quantitative methods to estimate G6PD activity in the Chittagong Hill Tracts, Bangladesh. PLoS ONE. 2017;12:e0169930.
- Weppelmann TA, von Fricken ME, Wilfong TD, Aguenza E, Philippe TT, Okech BA. Field trial of the CareStart Biosensor Analyzer for the determination of glucose-6-phosphate dehydrogenase activity in Haiti. Am J Trop Med Hyg. 2017;97:1262–70.
- WHO. Methods and techniques for clinical trials on antimalarial drug efficacy: genotyping to identify parasite populations. Geneva: World Health Organization; 2008.
- WHO. Methods for surveillance of antimalarial drug efficacy. Geneva: World Health Organization; 2009. http://whqlibdoc.who.int/publications/2009/9789241597531_eng.pdf. Accessed 13 Mar 2018.
- Warsame M, Hassan AM, Barrette A, Jibril AM, Elmi HH, Arale AM, et al. Treatment of uncomplicated malaria with artesunate plus sulfadoxinepyrimethamine is failing in Somalia: evidence from therapeutic efficacy studies and Pfdhfr and Pfdhps mutant alleles. Trop Med Int Health. 2015;20:510–7.
- Abreha T, Hwang J, Thriemer K, Tadesse Y, Girma S, Melaku Z, et al. Comparison of artemether-lumefantrine and chloroquine with and without primaquine for the treatment of *Plasmodium vivax* infection in Ethiopia: a randomized controlled trial. PLoS Med. 2017;14:e1002299.
- Kaplan EL, Meier P. Nonparametric estimation from incomplete observations. J Am Stat Assoc. 1958;53:457–81.
- WWARN. Clinical module: data management and statistical analysis plan. Oxford: WWARN; 2012.
- Schneider A, Hommel G, Blettner M. Linear regression analysis: part 14 of a series on evaluation of scientific publications. Dtsch Arztebl Int. 2010;107:776–82.
- Banoo S, Bell D, Bossuyt P, Herring A, Mabey D, Poole F, et al. Evaluation of diagnostic tests for infectious diseases: general principles. Nat Rev Microbiol. 2006;4(12 Suppl):S20–32.
- Ley B, Bancone G, von Seidlein L, Thriemer K, Richards JS, Domingo GJ, et al. Methods for the field evaluation of quantitative G6PD diagnostics: a review. Malar J. 2017;16:361.
- Mukhtar EA, Gadalla NB, El-Zaki SE, Mukhtar I, Mansour FA, Babiker A, et al. A comparative study on the efficacy of artesunate plus sulphadoxine/ pyrimethamine versus artemether-lumefantrine in eastern Sudan. Malar J. 2007;6:92.

- Adeel AA, Elnour FA, Elmardi KA, Abd-Elmajid MB, Elhelo MM, Ali MS, et al. High efficacy of artemether-lumefantrine and declining efficacy of artesunate + sulfadoxine-pyrimethamine against *Plasmodium falciparum* in Sudan (2010–2015): evidence from in vivo and molecular marker studies. Malar J. 2016;15:285.
- Gadalla NB, Abdallah TM, Atwal S, Sutherland CJ, Adam I. Selection of pfdhfr/pfdhps alleles and declining artesunate/sulphadoxine-pyrimethamine efficacy against *Plasmodium falciparum* eight years after deployment in eastern Sudan. Malar J. 2013;12:255.
- Mohamed AO, Abdel Hamid MM, Mohamed OS, Elkando NS, Suliman A, Adam MA, et al. Efficacies of DHA-PPQ and AS/SP in patients with uncomplicated *Plasmodium falciparum* malaria in an area of an unstable seasonal transmission in Sudan. Malar J. 2017;16:163.
- Warsame M, Hassan AH, Hassan AM, Arale AM, Jibril AM, Mohamud SA, et al. Efficacy of artesunate + sulphadoxine/pyrimethamine and artemether + lumefantrine and dhfr and dhps mutations in Somalia: evidence for updating the malaria treatment policy. Trop Med Int Health. 2017;22:415–22.
- Tjitra E, Baker J, Suprianto S, Cheng Q, Anstey NM. Therapeutic efficacies
 of artesunate-sulfadoxine-pyrimethamine and chloroquine-sulfadoxinepyrimethamine in vivax malaria pilot studies: relationship to *Plasmodium*vivax dhfr mutations. Antimicrob Agents Chemother. 2002;46:3947–53.
- Kolaczinski K, Durrani N, Rahim S, Rowland M. Sulfadoxine–pyrimethamine plus artesunate compared with chloroquine for the treatment of vivax malaria in areas co-endemic for *Plasmodium falciparum* and *P. vivax*: a randomised non-inferiority trial in eastern Afghanistan. Trans R Soc Trop Med Hyd. 2007;101:1081–7.
- Karunajeewa HA, Mueller I, Senn M, Lin E, Law I, Gomorrai PS, et al. A trial of combination antimalarial therapies in children from Papua New Guinea. N Engl J Med. 2008;359:2545–57.
- John GK, Douglas NM, von Seidlein L, Nosten F, Baird JK, White NJ, et al. Primaquine radical cure of *Plasmodium vivax*: a critical review of the literature. Malar J. 2012;11:280.
- 40. Grietens KP, Soto V, Erhart A, Ribera JM, Toomer E, Tenorio A, et al. Adherence to 7-day primaquine treatment for the radical cure of *P. vivax* in the Peruvian Amazon. Am J Trop Med Hyg. 2010;82:1017–23.
- 41. Khantikul N, Butraporn P, Kim HS, Leemingsawat S, Tempongko MA, Suwonkerd W. Adherence to antimalarial drug therapy among vivax malaria patients in northern Thailand. J Health Popul Nutr. 2009;27:4–13.
- 42. Pereira EA, Ishikawa EA, Fontes CJ. Adherence to *Plasmodium vivax* malaria treatment in the Brazilian Amazon Region. Malar J. 2011;10:355.
- Takeuchi R, Lawpoolsri S, Imwong M, Kobayashi J, Kaewkungwal J, Pukrittayakamee S, et al. Directly-observed therapy (DOT) for the radical 14-day primaquine treatment of *Plasmodium vivax* malaria on the Thai–Myanmar border. Malar J. 2010;9:308.

- Douglas NM, Poespoprodjo JR, Patriani D, Malloy MJ, Kenangalem E, Sugiarto P, et al. Unsupervised primaquine for the treatment of *Plasmodium vivax* malaria relapses in southern Papua: a hospital-based cohort study. PLoS Med. 2017;14:e1002379.
- 45. Leslie T, Rab MA, Ahmadzai H, Durrani N, Fayaz M, Kolaczinski J, et al. Compliance with 14-day primaquine therapy for radical cure of vivax malaria—a randomized placebo-controlled trial comparing unsupervised with supervised treatment. Trans R Soc Trop Med Hyg. 2004;98:168–73.
- 46. Ashley EA, Recht J, White NJ. Primaquine: the risks and the benefits. Malar J. 2014:13:418.
- Shekalaghe S, Drakeley C, Gosling R, Ndaro A, van Meegeren M, Enevold A, et al. Primaquine clears submicroscopic *Plasmodium falciparum* gametocytes that persist after treatment with sulphadoxine–pyrimethamine and artesunate. PLoS ONE. 2007;2:e1023.
- Howes RE, Piel FB, Patil AP, Nyangiri OA, Gething PW, Dewi M, et al. G6PD deficiency prevalence and estimates of affected populations in malaria endemic countries: a geostatistical model-based map. PLoS Med. 2012;9:e1001339.
- Louicharoen C, Patin E, Paul R, Nuchprayoon I, Witoonpanich B, Peerapittayamongkol C, et al. Positively selected G6PD-Mahidol mutation reduces *Plasmodium vivax* density in Southeast Asians. Science. 2009;326:1546–9.
- Pava Z, Burdam FH, Handayuni I, Trianty L, Utami RA, Tirta YK, et al. Submicroscopic and asymptomatic plasmodium parasitaemia associated with significant risk of anaemia in Papua, Indonesia. PLoS ONE. 2016;11:e0165340.
- Santana MS, Monteiro WM, Siqueira AM, Costa MF, Sampaio V, Lacerda MV, et al. Glucose-6-phosphate dehydrogenase deficient variants are associated with reduced susceptibility to malaria in the Brazilian Amazon. Trans R Soc Trop Med Hyg. 2013;107:301–6.
- Khim N, Benedet C, Kim S, Kheng S, Siv S, Leang R, et al. G6PD deficiency in *Plasmodium falciparum* and *Plasmodium vivax* malaria-infected Cambodian patients. Malar J. 2013;12:171.
- Kwiatkowski DP. How malaria has affected the human genome and what human genetics can teach us about malaria. Am J Hum Genet. 2005;77:171–92.
- Shah SS, Rockett KA, Jallow M, Sisay-Joof F, Bojang KA, Pinder M, et al. Heterogeneous alleles comprising G6PD deficiency trait in West Africa exert contrasting effects on two major clinical presentations of severe malaria. Malar J. 2016;15:13.

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