RESEARCH Open Access



Key to the females of Afrotropical *Anopheles* mosquitoes (Diptera: Culicidae)



Abstract

Background: In 1987, Gillies and Coetzee published a pictorial key for the morphological identification of adult female mosquitoes. Since then, several new species of anopheline mosquitoes have been described.

Methods: The 1987 key to adult female mosquitoes was used as the template for the current key.

Results: New species described in the literature over the past 32 years have been included. A list of all currently known Afrotropical species is provided. *Anopheles stephensi* is included for the first time as occurring on the African continent.

Conclusions: An updated key for the morphological identification of Afrotropical anopheline species is presented.

Keywords: Anopheles, Morphology, Identification, Key

Background

Dichotomous keys for the morphological identification of groups of organisms have been used for over 300 years. These keys lead the reader through a series of couplets, each one giving two choices of characters, until a species name is reached. For the anopheline mosquitoes of the Afrotropical Region, which includes some of the most efficient transmitters of malaria parasites in the world, the first key for their identification was published in 1931 [1], and the most recent printed version in 1987 [2], this last being a pictorial key containing line graphics of characters mentioned in each couplet. In the 32 years that have passed since Gillies and Coetzee published their key [2], several new species have been discovered, described and named.

Methods

The pictorial key to adult female anophelines in the Afrotropical Region [2] was used as the template for the current key. New illustrations were produced and new

*Correspondence: maureen.coetzee@wits.ac.za; maureenc@nicd.ac.za

¹ Wits Research Institute for Malaria, School of Pathology, Faculty of Health Sciences, University of the Witwatersrand, Johannesburg, South Africa Full list of author information is available at the end of the article

couplets inserted where appropriate to accommodate new species described since 1987.

Results

The present key is a revision of that presented in Gillies and Coetzee [2], with the addition of newly described species and the exclusion of subspecies. Table 1 provides a lists of species, authorship and current classification, while Table 2 gives the number of species described per decade since 1900. The user is encouraged to refer to both the 1968 volume of Gillies and De Meillon [3] and the 1987 supplement of Gillies and Coetzee for full species descriptions, biology and geographic distribution. More recent references include Sinka et al. [4], and Kyalo et al. [5].

The following species are not included in the keys: Anopheles ethiopicus lacks hindlegs [2], Anopheles erythraeus and Anopheles dualaensis adults are unknown [3, 6], and Anopheles eouzani lacks hindtarsomeres 4 and 5 [7]. The following new species are included: Anopheles okuensis [8] (Section I), Anopheles hervyi [6], Anopheles millecampsi [8] and Anopheles multicinctus [9] (Section IV), Anopheles rageaui [6] and Anopheles seretsei [10] (Section VII), Anopheles kosiensis [11] (Section IX),



© The Author(s) 2020. This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Coetzee *Malar J* (2020) 19:70 Page 2 of 20

Table 1 List of the species of *Anopheles* (excluding subspecies) in the Afrotropical Region, excluding Madagascar and associated islands

Subgenus	Series	Section	Group/complex*	Species	Authors
Anopheles	Myzorhynchus		Coustani	caliginosus	De Meillon, 1943
			Coustani	coustani	Laveran, 1900
			Coustani	crypticus	Coetzee, 1994
			Coustani	namibiensis	Coetzee, 1984
			Coustani	paludis	Theobald, 1900
			Coustani	symesi	Edwards, 1928
			Coustani	tenebrosus	Dönitz, 1902
			Coustani	ziemanni	Grünberg, 1902
				obscurus	(Grünberg, 1905)
	Anopheles			concolor	Edwards, 1938
ellia	Neomyzomyia	Smithii		caroni	Adam, 1961
	, , , , ,			faini	Leleup, 1952
				hamoni	Adam, 1962
				jebudensis	Froud, 1944
				lovettae	Evans, 1934
				rageaui	Mattingly & Adam, 1954
				smithii	Theobald, 1905
				vanhoofi	Wanson & Lebied, 1945
				wilsoni	Evans, 1934
		Ardensis		ardensis	
		Aruerisis			(Theobald, 1905)
				buxtoni	Service, 1958
				cinctus	(Newstead & Carter, 1910)
				dualaensis ,	Brunhes, Le Goff & Geoffroy, 1999
				deemingi , .	Service, 1970
				dureni	Edwards, 1938
				eouzani	Brunhes, Le Goff & Boussès, 2003
				kingi	Christophers, 1923
				machardyi	Edwards, 1930
				maliensis 	Bailly-Choumara & Adam, 1959
				millecampsi	Lips, 1960
				multicinctus	Edwards, 1930
				natalensis	(Hill & Haydon, 1907)
			Nili	carnevalei	Brunhes, Le Goff & Geoffroy, 1999
			Nili	nili	(Theobald, 1904)
			Nili	ovengensis	Awono-Ambene, Kengne, Simard, Antonio-Nkondjio & Fontenille, 2004
			Nili	somalicus	Rivola & Holstein, 1957
				vernus	Gillies & De Meillon, 1968
				vinckei	De Meillon, 1942
		Rhodesiensis		cameroni	De Meillon & Evans, 1935
				lounibosi	Gillies & Coetzee, 1987
				rhodesiensis	Theobald, 1901
				rodhaini	Leleup & Lips, 1950
				ruarinus	Edwards, 1940
	Myzomyia			azaniae	Bailly-Choumara, 1960
				barberellus	Evans, 1932
				bervoetsi	D'Haenens, 1961
				brunnipes	(Theobald, 1910)
				domicolus	Edwards, 1916

Coetzee *Malar J* (2020) 19:70 Page 3 of 20

Table 1 (continued)

ubgenus	Series	Section	Group/complex*	Species	Authors
				dthali	Patton, 1905
				erythraeus	Corradetti, 1939
				ethiopicus	Gillies & Coetzee, 1987
				flavicosta	Edwards, 1911
				fontinalis	Gillies & De Meillon, 1968
				gabonensis	Rahola, Makanga & Paupy, 2014
				moucheti	Evans, 1925
				schwetzi	Evans, 1934
				tchekedii	De Meillon & Leeson, 1940
				walravensi	Edwards, 1930
		Funestus	Funestus	aruni	Sobti, 1968
			Funestus	funestus	Giles, 1900
			Funestus	funestus-like	(See Spillings et al. [26])
			Funestus	parensis	Gillies, 1962
			Funestus	vaneedeni	Gillies & Coetzee, 1987
			Rivulorum	brucei	Service, 1960
			Rivulorum	fuscivenosus	Leeson, 1930
			Rivulorum	rivulorum	Leeson, 1935
			Rivulorum	rivulorum-like	(see Cohuet, et al. [27])
			Tilvaloram	confusus	Evans & Leeson, 1935
				culicifacies	Giles, 1901
				leesoni	Evans, 1931
				longipalpis	(Theobald, 1903)
		Marshallii-Hancocki		austenii	(Theobald, 1905)
		Maistrailli-Francocki		berghei	Vincke & Leleup, 1949
				brohieri	Edwards, 1929
				gibbinsi hancocki	Evans, 1935
					Edwards, 1929
				hargreavesi	Evans, 1927
			A A - male - III	harperi	Evans, 1936
			Marshallii	hughi	Lambert & Coetzee, 1982
			Marshallii	kosiensis	Coetzee, Segerman & Hunt, 1987
			Marshallii	letabensis	Lambert & Coetzee, 1982
			Marshallii	marshallii 	(Theobald, 1903)
				mortiauxi	Edwards, 1938
				mousinhoi · , · ·	De Meillon & Pereira, 1940
				njombiensis	Peters, 1955
				seydeli	Edwards, 1929
		Wellcomei		distinctus	(Newstead & Carter, 1911)
				erepens	Gillies, 1958
				theileri	Edwards, 1912
		6 41 -		wellcomei	Theobald, 1904
		Demeilloni		carteri	Evans & De Meillon, 1933
				demeilloni	Evans, 1933
				freetownensis	Evans, 1925
				garnhami	Edwards, 1930
				keniensis	Evans, 1931
				lloreti	Gil Collado, 1935
				sergentii	(Theobald, 1907)

Coetzee Malar J (2020) 19:70 Page 4 of 20

Table 1 (continued)

Subgenus	Series	Section	Group/complex*	Species	Authors
	Pyretophorus			christyi	(Newstead & Carter, 1911)
				daudi	Coluzzi, 1958
			Gambiae	amharicus	Hunt, Wilkerson & Coetzee, 2013
			Gambiae	arabiensis	Patton, 1905
			Gambiae	bwambae	White, 1985
			Gambiae	coluzzii	Coetzee & Wilkerson, 2013
			Gambiae	fontenillei	Barrón, Paupy, Rahola, Akone-Ella, Ngangue, Wilson-Bahun, Pombi, Kengne, Costantini, Simard, González & Ayala, 2019
			Gambiae	gambiae	Giles, 1902
			Gambiae	quadriannulatus	(Theobald, 1911)
			Gambiae	melas	Theobald, 1903
			Gambiae	merus	Donitz, 1902
	Paramyzomyia			azevedoi	Ribeiro, 1969
				cinereus	Theobald, 1901
				listeri	De Meillon, 1931
				multicolor	Cambouliu, 1902
				seretsei	Abdulla-Khan, Coetzee & Hunt, 1998
				turkhudi	Liston, 1901
	Neocellia			dancalicus	Corradetti, 1939
				hervyi	Brunhes, Le Goff & Geoffroy, 1999
				maculipalpis	Giles, 1902
				pretoriensis	(Theobald, 1903)
				rufipes	(Gough, 1910)
				salbaii	Maffi & Coluzzi, 1958
				stephensi	Liston, 1901
	Cellia			argenteolobatus	(Gough, 1910)
				brumpti	Hamon & Rickenbach, 1955
				cristipalpis	Service, 1977
				cydippis	De Meillon, 1931
				murphyi	Gillies & De Meillon, 1968
				pharoensis	Theobald, 1901
				squamosus	Theobald, 1901
				swahilicus	Gillies, 1964
Christya**				implexus	(Theobald, 1903)
				okuensis	Brunhes, Le Goff & Geoffroy, 1997

^{*}Anopheles gambiae, An. nili and An. marshallii are referred to as complexes, while An. coustani, An. funestus and An. rivulorum are groups

Anopheles gabonensis [12] (Section X), and Anopheles carnevalei [6] and Anopheles ovengensis [13] (Section XI).

Discussion

A major addition to the key is the inclusion of *Anopheles stephensi* (Section IV), the Asian malaria vector with distribution from the Middle East to China. This

species was first detected on the African continent in Djibouti in September 2012 and subsequently in February 2013 [14] through to December 2017 [15]. It has also recently been found in Ethiopia in 2016 [16]. The species is similar to those belonging to the *Anopheles gambiae* complex—mosquitoes with speckled legs—but differs by having the wing with two pale spots in the 2nd main dark area of the costa and vein 1, thus being

^{**}Previously a Series in the subgenus Anopheles, Christya was elevated to subgeneric status by Harbach and Kitching in 2016 [28]

Coetzee Malar J (2020) 19:70 Page 5 of 20

Table 2 Number of species of Afrotropical Anopheles described per decade since 1900

Decade	No. species	Author/co-author (number of species authored/co-authored)
1900–1910	35	Theobald (18), Giles (3), Donitz (2), Gough (2), Grunberg (2), Liston (2), Patton (2), Cambouliu (1), Carter (1), Haydon (1), Hill (1), Laveran (1), Newstead (1)
1911-1920	6	Edwards (3), Carter (2), Newstead (2), Theobald (1)
1921-1930	13	Edwards (9), Evans (2), Christophers (1), Leeson (1)
1931-1940	24	Evans (12), De Meillon (6), Edwards (4), Leeson (3), Corradetti (2), Gil Collado (1), Pereira (1)
1941-1950	6	De Meillon (2), Leleup (2), Froud (1), Lebied (1), Lips (1), Vincke (1), Wanson (1)
1951–1960	13	Adam (2), Bailly-Choumara (2), Coluzzi (2), Service (2), Gillies (1), Hamon (1), Holstein (1), Leleup (1), Maffi (1), Mattingly (1), Peters (1), Rickenbach (1), Rivola (1)
1961-1970	11	Gillies (5), De Meillon (3), Adam (2), D'Haenens (1), Ribeiro (1), Service (1), Sobti (1)
1971-1980	1	Service (1)
1981-1990	8	Coetzee (7), Gillies (3), Lambert (2), Hunt (1), Segerman (1), White (1)
1991-2000	6	Brunhes (4), Le Goff (4), Geoffroy (4), Coetzee (2), Abdulla-Khan (1), Hunt (1)
2001–2010	4	Coetzee (2), Fontenille (2), Kengne (2), Simard (2), Antonio-Nkondjio (1), Awono-Ambene (1), Bousses (1), Brooke (1), Brunhes (1), Chiphwanya (1), Cohuet (1), Hunt (1), Koekemoer (1), Le Goff (1), Spillings (1), Toto (1)
2011–2020	4	Coetzee (2), Paupy (2), Rahola (2), Wilkerson (2), Akone-Ella (1), Ayala (1), Barrón (1), Costantini (1), González (1), Hunt (1), Kengne (1), Makanga (1), Ngangue (1), Pombi (1), Simard (1), Wilson-Bahun (1)

similar to *Anopheles maculipalpis* and *Anopheles pretoriensis*, from which it differs by not having hindtarsomeres 4 and 5 all pale.

There are several groups of species where morphological identification is not possible using only the adult females, either because the adults look identical or because of overlap in morphological variation. Some of these species can be identified on immature characters, thus requiring eggs or larvae [3], while others require genetical methods, such as chromosomal inversions [17] or molecular assays [18, 19]. Such groups include:

- (a) the well-known Anopheles gambiae complex (An. gambiae, Anopheles coluzzii [20], Anopheles arabiensis, Anopheles quadriannulatus, Anopheles amharicus [20], Anopheles fontenillei [21], Anopheles bwambae, Anopheles melas, Anopheles merus);
- (b) the Anopheles funestus group (Anopheles funestus, An. funestus-like, Anopheles parensis, Anopheles vaneedeni, Anopheles aruni, Anopheles confusus, Anopheles leesoni, Anopheles rivulorum, An. rivulorum-like, Anopheles brucei, Anopheles fuscivenosus);
- (c) the Anopheles nili complex (An. nili, Anopheles somalicus);
- (d) the Anopheles marshallii complex (An. marshallii, Anopheles letabensis, Anopheles hughi, Anoph-

- eles kosiensis) and its allies Anopheles hargreavesi, Anopheles gibbinsi and Anopheles mousinhoi;
- (e) Anopheles squamosus/cydippis, the former of which is known to consist of at least five chromosomal forms (Green and Hunt, unpublished data);
- (f) Anopheles coustani/crypticus/namibiensis in southern Africa.

The definition of "complex", as applied to the genus *Anopheles*, is a group of species that are virtually morphologically identical but are otherwise considered valid species. The use of the term "group" denotes species that are morphologically very similar at the adult stage but many can be distinguished at the immature stages.

Except for those of medical importance, the above list is just a small sample of species groups about which we know very little biologically in terms of feeding/resting preferences or their role in malaria parasite transmission. Basic taxonomic research, aligned with molecular analyses, is still very much needed in the Culicidae.

Conclusions

An updated key for the morphological identification of Afrotropical anopheline species is presented. This key should be used in conjunction with earlier works giving full species descriptions, biology, medical importance and distribution.

Coetzee Malar J (2020) 19:70 Page 6 of 20

Table 3 The terminology used for wing venation since 1938 [25] is given

	Evans [25]	Gillies & De Meillon [3]; Gillies & Coetzee [2]	Snodgrass [29]	Harbach & Knight [22]	McAlpine [30]	Cumming & Wood [24]
Costa	Costa	Costa	Costa	Costa	Costa	Costa
Sub-costa	Sub-costa	Sub-costa	Sub-costa	Sub-costa	Sub-costa	Sub-costa
1	1st vein	1st vein	R ₁ —1st vein	R ₁ —radius-one	R ₁ —anterior radius	R ₁ —anterior radius
2	2.1—2nd vein upper branch 2.2—2nd vein lower branch	2nd vein upper branch 2nd vein lower branch	R ₂ —2nd vein upper branch R ₃ —2nd vein lower branch	R ₂ —radius-two R ₃ —radius-three R ₂₊₃ —stem of vein	R ₂ —upper branch of 2nd radius R ₃ —lower branch of 2nd radius R ₂₊₃ —2nd branch of radius	R ₂ —upper branch of 2nd radius R ₃ —lower branch of 2nd radius R ₂₊₃ —2nd branch of radius
3	3rd vein	3rd vein	R ₄₊₅ —3rd vein	R ₄₊₅ —radius-four- plus-five	R ₄₊₅ —3rd branch of radius	R ₄₊₅ —3rd branch of radius
4	4.1—4th vein upper branch 4.2—4th vein lower branch	4th vein upper branch 4th vein lower branch	M ₁₊₂ —4th vein upper branch M ₃ —4th vein lower branch M—stem of vein	M_1 —media-one M_2 —media-two M_{1+2} —stem of vein	M ₁ —1st branch of media M ₂ —2nd branch of media	M ₁ —1st branch of media M ₂ —2nd branch of media
5	5.1—5th vein upper branch 5.2—5th vein lower branch	5th vein upper branch 5th vein lower branch	Cu ₁ —5th vein upper branch Cu ₂ —5th vein lower branch Cu—stem of vein	M ₃₊₄ —media-three- plus-four CuA—cubitus anterior M—stem of vein	CuA ₁ —1st branch of anterior cubital CuA ₂ —2nd branch of anterior cubital	M ₃₊₄ —3rd +4th branch of media CuA—anterior cubital
6	6th vein	6th vein	A—anal vein	1A—anal vein	A ₁ —anal vein	CuP—posterior cubital

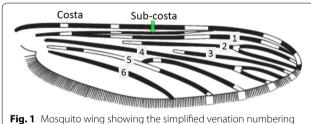


Fig. 1 Mosquito wing showing the simplified venation numbering system of Evans [25]

The key layout

Characters are presented in 'couplets' where two options are presented, giving two different outcomes, eventually ending at a species name. The illustration(s) for the first option of each couplet is on the left (or rarely, in the centre) and for the second option on the

right. General terminology follows that of Harbach and Knight [22, 23].

Terminology of the wing venation has changed over the past 80 years in attempts to align Culicidae with the rest of the Diptera Family (Table 3). The terminology proposed in the recent Manual of Afrotropical Diptera [24] has been challenged by culicid taxonomists (manuscript reviewers), specifically around the terms used for the posterior veins (veins 5 and 6 in Fig. 1). Since consensus on the terminology has not yet been reached, and given that the malaria vector control field workers in Africa have been using the *Anopheles* identification keys published 32 years ago [2] that use the numbering shown in Fig. 1, this simplified system is used here. It avoids unnecessary repetition of the various terms in each couplet and reference can be made to Table 3 where recent terminology is required.

Coetzee *Malar J* (2020) 19:70 Page 7 of 20

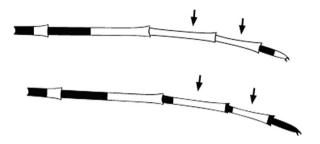
Key 1	to a	dult	: fem	ales
-------	------	------	-------	------

l. -	Abdominal segments with laterally projecting tufts of scales on segments II–VIISection I Abdominal segments not so2
THE RELL	
2.	Hindtarsus with at least last 2 hindtarsomeres entirely pale
-	
3. -	Hindtarsomere 5 mainly or entirely dark, hindtarsomere 4 white
	1.
	Legs speckled, sometimes sparsely Section IV Legs not speckled5
_	
_	Wing entirely dark or with pale spots confined to costa and vein 1Section V
5.	Wing entirely dark or with pale spots confined to costa and vein 1Section V
5.	Wing entirely dark or with pale spots confined to costa and vein 1

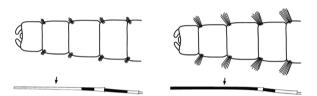
8. Maxillary palpus with 4 pale bands Section VIII
- Maxillary palpus with less than 4 pale bands9
The state of the s
9. Wing with pale interruption in 3rd main dark area
(preapical dark spot) of vein 1, sometimes fused with
preceding pale area Section IX
 3rd main dark area without pale interruption10
10 W
10. Wing with 2 pale spots on upper branch of vein 5
Section X
- Wing with I pale spot on upper branch of vein 5
Section XI
'a atian I Managaria a a sith latanalla mania atian a afta
Section I. Mosquitoes with laterally projecting tufts
of abdominal scales
of abdominal scales
· · · · · · · · · · · · · · · · · · ·
of abdominal scales
of abdominal scales 1. Wing almost entirely dark, costa without pale spotsbrumpti
1. Wing almost entirely dark, costa without pale spots
of abdominal scales 1. Wing almost entirely dark, costa without pale spotsbrumpti
1. Wing almost entirely dark, costa without pale spots
1. Wing almost entirely dark, costa without pale spots
1. Wing almost entirely dark, costa without pale spots
1. Wing almost entirely dark, costa without pale spots
1. Wing almost entirely dark, costa without pale spots
1. Wing almost entirely dark, costa without pale spots
1. Wing almost entirely dark, costa without pale spots
1. Wing almost entirely dark, costa without pale spotsbrumpti - Wing with abundant pale areas, costa with at least 4 pale spots2 2 (1). Hindtarsomeres 1 to 5 entirely darkargenteolobatus
1. Wing almost entirely dark, costa without pale spots
1. Wing almost entirely dark, costa without pale spotsbrumpti - Wing with abundant pale areas, costa with at least 4 pale spots2 2 (1). Hindtarsomeres 1 to 5 entirely darkargenteolobatus
1. Wing almost entirely dark, costa without pale spotsbrumpti - Wing with abundant pale areas, costa with at least 4 pale spots2 2 (1). Hindtarsomeres 1 to 5 entirely darkargenteolobatus (southern Africa) murphyi
1. Wing almost entirely dark, costa without pale spots
1. Wing almost entirely dark, costa without pale spots
1. Wing almost entirely dark, costa without pale spots
1. Wing almost entirely dark, costa without pale spots
1. Wing almost entirely dark, costa without pale spots
1. Wing almost entirely dark, costa without pale spots
1. Wing almost entirely dark, costa without pale spots
1. Wing almost entirely dark, costa without pale spots
1. Wing almost entirely dark, costa without pale spots
1. Wing almost entirely dark, costa without pale spots
1. Wing almost entirely dark, costa without pale spots
1. Wing almost entirely dark, costa without pale spots
1. Wing almost entirely dark, costa without pale spots

Coetzee Malar J (2020) 19:70 Page 8 of 20

4(3).	Hindtars	someres	3	and	4	all	white	or	narro	wly
dark	k basally, !	5 all dark	O	r at le	eas	t ba	sal 0.5	dar	k	5
	4.									_



- 5 (4). Moderate-sized species; abdominal scale-tufts short and dark; 0.5 or more of hindtarsomere 1 palecristipalpis



- No pale fringe spot opposite lower branch of vein 5implexus

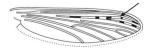




- Hindtarsomere 5 all dark and 4 with much less than apical 0.5 pale8



- 8 (7). Very small species (wing length 2.5–2.8 mm); wing with upper branch of vein 2 largely pale swahilicus





Section II. Mosquitoes with hindtarsomeres 4 and 5 entirely white; abdomen without laterally projecting tufts of scales

- 3 (2). Maxillary palpus with 3 pale bands, usually with some speckling; vein 1 of wing with 2 pale spots in 2nd main dark area (median dark spot) maculipalpis



- 4 (3). Midtarsomeres 2 to 4 entirely dark; vein 1 of wing dark at base, basal 0.5 of stem of vein 4 with small pale areasmaliensis
- Midtarsomeres 2 to 4 with pale apices; vein 1 of wing pale at base, basal 0.5 of stem of vein 4 entirely pale
 deemingi





- 5 (2). Hindtarsomere 1 broadly pale at apex; vein 1 of wing with 2 pale spots in 2nd main dark areapretoriensis
- Hindtarsomere 1 narrowly pale or dark at apex; vein
 1 of wing with 1 pale spot in 2nd main dark area6

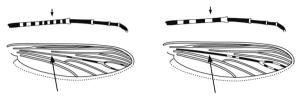




Coetzee Malar J (2020) 19:70 Page 9 of 20

6 (5). Foretarsomere 1 with 5–9 pale rings; stem of vein 4 of wing largely palemachardyi

 Foretarsomere 1 with 2-4 pale rings; stem of vein 4 of wing largely dark7



7 (6). Fore- and midtarsomeres 2 and 3 pale at apex; wing with fringe spot opposite vein 6 natalensis

- Fore- and midtarsomeres 2 and 3 dark apically; no fringe spot opposite vein 6buxtoni



8 (1). Maxillary palpus very shaggy and unbanded or with 1-4 irregular narrow pale bands9

- Maxillary palpus smooth with 3 pale bands, the 2 distal ones broad or rarely fused14



9 (8). Maxillary palpus without pale bands; no pale spot at apex of hindtibia or base of hindtarsomere 1caliginosus

 Maxillary palpus with 1-4 pale bands; apex of hindtibia broadly or narrowly pale10

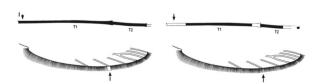


10 (9). Hindtarsomere 3 entirely pale11 - Hindtarsomere 3 dark at base12



11 (10). Base of hindtarsomere 1 dark; pale fringe spot present opposite lower branch of wing vein 5 paludis

- Base of hindtarsomere 1 broadly pale; no pale fringe spot opposite lower branch of wing vein 5 ... coustani (in part)



12 (10). Hind tarsomere 1 entirely dark basally or at most with a very narrow band of pale scales not as broad as the width of the tarsomeretenebrosus (in part)

- Hind tarsomere 1 broadly pale at base, pale area at least as long as broad13



13 (12). Apex of hindtibia with a pale streak 3–5 times as long as broad; apical pale band on hindtarsomere 2 0.13-0.4 length of tarsomerecoustani (in part)

> crypticus (S. Africa only)

Pale streak on hindtibia 1–3 times as long as broad; apical pale band on hindtarsomere 2 narrow, 0.07-0.13 length of segmentziemanni namibiensis



14 (8). 3rd main dark area on wing vein 1 without a pale interruption; foretarsomeres 1 to 3 usually without distinct apical pale bandsrufipes

- 3rd main dark area on wing vein 1 with a pale interruption, or with a short extension of the subcostal pale spot into the dark area on vein 1; foretarsomeres 1 to 3 with apical pale bands15





15 (14). Hindtarsomere 3 entirely pale......hancocki brohieri

(in part)

Hindtarsomere 3 not so......brohieri (in part) W. Africa

theileri mainly E. & S. Africa



Coetzee Malar J (2020) 19:70 Page 10 of 20

Section III. Mosquitoes with hind tarsomere 5 mainly or entirely dark, tarsomere 4 white; abdominal segments without laterally projecting tufts of scales

1.	Femora and	tibiae	speckled		kingi
----	------------	--------	----------	--	-------

- Femora and tibiae with at most apical bands only2



- 2 (1). Maxillary palpus shaggy; costa and vein 1 of wing without usual main dark areassymesi



Section IV. Mosquitoes with speckled legs, hindtarsomeres 4 and 5 not entirely pale; abdominal segments without laterally projecting tufts of scales

- 1. Maxillary palpus with 3 pale bands 2
- Maxillary palpus with 4 pale bands6



- 2 (1). Maxillary palpus with apical 2 pale bands very broad, speckling on palpus segment 3; 2nd main dark area on wing vein 1 with 2 pale interruptions *stephensi*





- 5 (4). Foretarsomere 1 with some speckling; base of costa with 2 pale spots; stem of wing vein 2 entirely palesalbaii



- 6 (1). All tarsi completely dark; wing without pale fringe spots posterior to vein 3*vernus* (in part)





Coetzee *Malar J* (2020) 19:70 Page 11 of 20

7 (6). 3rd main dark area of vein 1 with a pale interruption, sometimes fused with preceding pale area	Section V. Mosquitoes with wings entirely dark or with pale spots confined to costa and vein 1; legs not speckled, hindtarsomeres 4 and 5 not entirely pale; abdominal segments without laterally projecting tufts of scales
- 3rd main dark area without pale interruption8	 Wings entirely dark. or unicolorous
8 (7).Hindtarsomere 2 with about apical 0.4 to 0.5 white and the rest dark	
	2 (1).Maxillary palpus with 2 well-marked pale bands hindfemur and hindtibia narrowly pale at apexconcolor
	Maxillary palpus and legs entirely dark3
 9 (8).Hindtarsomeres 2 to 4 with apical pale rings and otherwise dark except for 1 to 2 pale spots; no pale fringe spot opposite wing vein 6ardensis Hindtarsomeres 2 to 4 with conspicuous dark and pale rings in addition to apical pale bands; pale fringe spot present opposite vein 6	3 (2).Large species, wing length 4 mm or more <i>ruarinus</i>
	 Small species, wing length 3.5 mm or less
	(in part) – General coloration dark brown, scutum not so; cave- dwelling
10 (9).Foretarsomeres mainly pale with narrow dark markings	(in part) 5 (1).Maxillary palpus with 2 to 3 pale bands, pale at apex (sometimes indistinct)
 11 (10). Scales on abdominal tergum VIII dense and distributed over whole tergum, sometimes with a few scales on lateral borders of tergum VII	6 (5).Erect head scales narrow, rod-like, all scales yellowish throughout; semi-arid regions onlydthal. – Erect head scales broader, scales white on vertex dark laterally; all regions

Coetzee *Malar J* (2020) 19:70 Page 12 of 20

7 (6).Pale and dark areas on wing poorly contrasted; semi-arid regions onlyrhodesiensis (in part) Pale and dark areas on wing well contrasted8	Section VI. Mosquitoes without a pale spot on basal 0.5 of costa; pale spots not confined to costa and vein 1; legs not speckled, hindtarsomeres 4 and 5 not entirely pale; abdomen without projecting tufts of scales
	Maxillary palpus shaggy to near tip2 – Maxillary palpus smooth except at extreme base3
8 (7). Pale areas on wing very narrow, subcostal pale spot present on costa only; cave-dwellingrodhaini – Pale areas on wing broader, subcostal pale spot on costa and vein 1rhodesiensis (in part) lounibosi	2 (1). Maxillary palpus entirely dark; hindtarsomeres 3 and 4 dark or narrowly pale at apicesobscurus (in part) – Maxillary palpus with pale scales forming more or less definite pale bands; hindtarsomeres 3 and 4 narrowly or broadly pale at apicestenebrosus (in part)
9 (5). Maxillary palpus with 3 pale bands, dark at apex	
10 (9). Cave-dwelling species; colour and contrast of dark and pale areas on wing variable	3 (1). Maxillary palpus with apex dark, sometimes only narrowly so
 Erect head scales broader, scales white on vertex, dark laterallyrhodesiensis	smithi (in part)

Coetzee Malar J (2020) 19:70 Page 13 of 20



- 6 (5). Maxillary palpus with 3 pale bands, subapical band broad and about equal in length to apical band .7
- Maxillary palpus either with 4 pale bands or if with 3 bands then subapical band much shorter than the apical band9



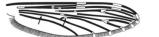
- 7 (6). Wing, apart from costa, generally very pale, basal 0.5 of stems of veins 2 and 4 entirely palewellcomei (in part)
- Dark areas on wing greater than or about equal to pale areas, basal 0.5 of stems of veins 2 and 4 largely dark8



- Pale fringe spots present opposite all veins from wing apex to vein 5, stem of vein 5 broadly dark near base
 keniensis
 (in part)

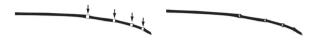


- 9 (6). Wing vein 5 entirely dark except for a single pale spot on the upper branchfuscivenosus (in part)
- Wing vein 5 with extensive pale areas, upper branch of vein 5 with 2 pale spots10





- Hindtarsomeres 1 to 4 entirely dark or with a few pale scales at apices of 1 to 3; scutum scales broad11



- 11 (10). Median scutal scales yellowish or bronze, white elsewhereschwetzi (in part)
- Scutal scales white throughoutwalravensi
 (in part)
 schwetzi
 (in part)

Section VII. Mosquitoes with maxillary palpus dark at apex or without distinct apical pale band; at least 1 pale spot on basal 0.5 of costa, pale scales not confined to costa and wing vein 1; legs not speckled, hindtarsomeres 4 and 5 not entirely pale; abdomen without laterally projecting tufts of scales

- 1. Maxillary palpus entirely dark or without distinct pale bands2
- Maxillary palpus with 3 pale bands5



- Wing with well-contrasting pale and dark areas, basal 0.25 of costa with at least 1 pale area, even if narrow; head scales not so



Coetzee *Malar J* (2020) 19:70 Page 14 of 20

 3 (2). Costa with humeral pale spot, no subapical (preapical) pale spot on costa and vein 1obscurus (in part) Costa without a humeral pale spot, subapical pale spot present on costa and vein 1	8 (6). Wing, apart from costa and vein 1, predominantly dark, no pale spots on basal 0.25 of costa9 Pale and dark areas on wing about equally distributed, humeral and presector pale spots present on costa
4 (3). Wing with pale fringe spots opposite all veins except vein 6jebudensis - Wing with no pale fringe spots posterior to vein 3faini	9 (8). Wing vein 6 darkrageaui — Wing vein 6 with proximal pale spot10
5 (1). Wing generally pale, contrast between pale and dark areas, apart from costa and vein 1, poorly defined	 10 (9). Basal 0.2 of wing vein 1 either dark or with a proximal pale patch not extending to basesmithii (in part) Basal 0.2 of wing vein 1 entirely pale
6 (5). 2nd main dark area of wing vein 1 with 2 pale interruptions	11 (10). Wings scantily scaled, all wing scales very narrow
7 (6).Pale bands on maxillary palpus very narrow, at apices of segments 2 to 4 and not overlapping the joints; upper branch of wing vein 5 with a single pale spotwilsoni (in part)	
Pale bands on maxillary palpus variable in width, distal 2 bands overlapping the joints; upper branch of wing vein 5 with 2 pale spotsrufipes (in part)	12 (8). Basal pale band of maxillary palpus about equal to or slightly shorter than median band, broadly overlapping base of 3rd segment
	shorter than median band, scarcely overlapping base of 3rd segment, or both basal and median pale bands very narrow

Coetzee Malar J (2020) 19:70 Page 15 of 20

13 (12). Base of costa pale*multicolor* (north-east Africa only)

- Base of costa dark ...listeri

(southern Africa only)

azevedoi

(south-western Africa only)

seretsei

(Botswana only)

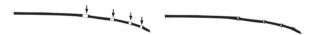


Section VIII. Mosquitoes with smooth, 4-banded maxillary palpus, pale at apex; at least 1 pale spot on basal 0.5 of costa, pale scales not confined to costa and wing vein 1; legs not speckled, hindtarsomeres 4 and 5 not entirely pale; abdomen without laterally projecting tufts of scales

- 3rd main dark area without a pale interruption.......3



- Abdominal terga without such scales; hindtarsomeres entirely dark or with a few pale scales at apices of hindtarsomeres 1 to 3schwetzi (in part)



- 3 (1). 2nd main dark area of wing vein 1 with 2 pale interruptions......wilsoni (in part)
- 2nd main dark area of wing vein 1 with 1 pale interruption4





- 5 (4). No pale fringe spots on wing posterior to vein 3; femora and tibiae inconspicuously speckled*vernus* (in part)
- Pale fringe spots on wing present opposite veins posterior to vein 3, sometimes including vein 6; femora and tibiae not speckled.



- 6 (5). Stem of wing vein 5 pale, at and adjacent to the forkgarnhami (in part)
- Fork of wing vein 5 dark7





- 7(6). Wing length 4 mm or less; decumbent scutal scales not extending onto scutellumdemeilloni (Berg River form)
- Wing length 4.4 mm or more; some decumbent scales present on scutellum as well as scutum .carteri (in part)

Section IX. Mosquitoes with a pale interruption in 3rd main dark area (preapical dark spot) of wing vein 1 or this area entirely pale; at least 1 pale spot on basal 0.5 of costa, pale scales not confined to costa and vein 1; maxillary palpus with 3 pale bands, pale at apex; legs not speckled, hindtarsomeres 4 and 5 not entirely pale; abdomen without laterally projecting tufts of scales

- 1. 2nd and 3rd main dark areas of wing (median and preapical dark spots) absent from vein 1wellcomei (in part)
- 2nd and 3rd main dark areas present on vein 12



- 2 (1). Hindtarsomere 5 entirely pale, hindtarsomere 4 with broad apical and basal pale bandsseydeli
- Hindtarsomere 5 entirely dark, hindtarsomere 4 with narrow apical and basal pale bands......3



Coetzee Malar J (2020) 19:70 Page 16 of 20

3 (2). Upper branch of wing vein 5 with 1 pale spot, sometimes a vestigial 2nd pale spot4 - Upper branch of wing vein 5 with 2 well-developed pale spots5
 4 (3). Pale fringe spot present opposite wing vein 6; foretarsomeres 1 to 4 with conspicuous basal and apical pale bands
 5 (3). Subapical pale band on maxillary palpus very narrow, confined to apex of 3rd segment
6 (5). Wing with base of costa with 2 pale interruptions
+
7 (5). Hindtarsomeres either all dark or with pale bands on tarsomeres 1 and 2 only

8 (7). Scutal fossae and lateral areas of scutum above wing root (supraalar area) without scalesharperi Scutal fossae and lateral areas of scutum above wing root (supraalar area) with scattered or abundant broadish scales9 9 (8). Subapical pale band on maxillary palpus about equal to or slightly shorter than apical bandnjombiensis - Subapical pale band on maxillary palpus much narrower than apical bandwalravensi (in part) 10 (7). Apical pale bands on hindtarsomeres 1 to 4 very broad, at least twice the apical width of the tarsomeres......austenii - Hindtarsomeres 1 to 4 with narrow pale bands, as long or shorter than the width of the tarsomeres11 11 (10). Wing vein 3 largely dark or broadly dark at either end; scutal scales very narrow and goldengibbinsi - Wing vein 3 narrowly dark at ends; scutal scales various12 12 (11). Scutal scales as in Ahargreavesi Scutal scales as in Bmousinhoi Scutal scales as in Cmarshallii letabensis kosiensis hughi Scutal scales as in Dgibbinsi (in part) a d C

Coetzee Malar J (2020) 19:70 Page 17 of 20

Section X. Mosquitoes with upper branch of wing vein 5 with 2 pale spots, no pale interruptions in 3rd main dark area of vein 1, at least 1 pale spot on basal half of costa, pale scales not confined to costa and vein 1; maxillary palpus with 3 pale bands or less, pale at apex; legs not speckled, hindtarsomeres 4 and 5 not entirely pale; abdomen without laterally projecting tufts of scales

1.	Maxillary palpus with only apical pale bandgabonensis
	Maxillary palpus with 3 pale bands2

flame.	Manuel		
	" "		

2 (1).Subapical	pale	band	on	maxillar	y palpus	s broad,
about equal to	or lo	onger t	han	apical da	rk band	3







- Base of costa with 2 pale interruptions, 3rd main dark area equal to or narrower than subcostal pale spot6

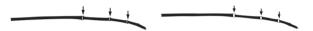


6 (5). Pale fringe spot present opposite wing vein 6
......lloreti

- No pale fringe spot opposite vein 6barberellus



- 7 (3). Apices of hindtarsomeres 3 and 4 dark or at most with a few pale scales8



- 8 (7). Base of costa with 1 or no pale interruption9

 Base of costa with 2 pale interruptions10
- 9 (8). Wing vein 6 either with pale fringe spot or with pale scales at apex of veinbrucei
- Wing vein 6 without pale fringe spot and no pale scales at apexrivulorum (in part)



- 10 (8). Scutal scales fairly broad, extending over whole scutum and onto scutellumcarteri (in part)
- Scutal scales variable, but decumbent scales confined to at most anterior 0.66 of scutum11

- 12 (11). Hindtarsomeres entirely dark; preaccessory dark spot on wing vein 1 usually absent freetownensis



Coetzee *Malar J* (2020) 19:70 Page 18 of 20

13 (7). 3rd main dark area of costa equal to or shorter than subapical pale spot	18 (17). Wing with fork of vein 5 palegarnhami (in part)
(in part) - 3rd main dark area much longer than subapical pale spot14	- Wing with fork of vein 5 dark19
	19 (18). Base of costa with 2 pale interruptionsdemeilloni
14 (3). Scutal scales broadish and white, only slightly less dense on posterior 0.33 of scutum than anteriorly, and extending onto scutellum	(in part) - Base of costa with 1 or no pale interruption20
(in part) - Scutal scales on posterior 0.33 of scutum scanty, nar-	
row and yellowish-brown15	20 (19). Small species, wing length about 2.4–3.3 mm
	(in part) - Small or moderate-sized species, wing length 2.9– 4.2 mmdemeilloni (in part)
	Section XI. Mosquitoes with upper branch of wing vein 5 with 1 pale spot, no pale interruptions on 3rd main dark area of vein 1, at least 1 pale spot on basal 0.5 of costa, pale scales not confined to costa and vein 1; maxillary palpus with 3 pale bands or less, pale at apex; legs
15 (14). Moderate-sized species, wing length more than 3.2 mmkeniensis	not speckled, hindtarsomeres 4 and 5 not entirely pale; abdomen without laterally projecting tufts of scales
(in part) - Small species, wing length 3.0 mm or less	1. Maxillary palpus with only apex pale2 Maxillary palpus with 3 pale bands5
vein 6	
wing with thinge spot opposite vein o	2 (1). Base of costa with large (presector) pale spot, base
	of vein 1 pale3 Base of costa dark or with small pale spot, base of vein 1 dark4
17 (2). 3rd main dark area of costa equal to or shorter	
than subapical pale spot	2 (2) I assemble of suite a series 2 and a series and 1 of
- 3rd main dark area of costa much longer than subapical pale spot18	3 (2). Lower branch of wing vein 2 and upper branch of vein 4 with distinct pale spots

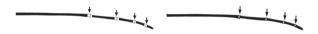
Coetzee *Malar J* (2020) 19:70 Page 19 of 20

Congo form

 Subapical pale spot shorter, usually much shorter, than apical dark spot, no pale fringe spot opposite upper branch of vein 5nili



- Pale banding on hindtarsomeres narrow and apical only6



- 6 (5). Preaccessory dark spot on wing vein 1 about twice as long as pale spot on either side of it *fuscivenosus* (in part)
- Preaccessory dark spot absent or, if present, shorter or only slightly longer than adjoining pale spots7



- 7 (6). Basal area of wing vein 1 proximal to 1st main dark area, pale with a broad dark spotculicifacies
- Basal area of wing vein 1 entirely pale.....8





- Subapical pale band on maxillary palpus much shorter than apical dark band, OR 3rd main dark area longer than subapical pale spot......9





(extreme S. Africa only)

Acknowledgements

Dr. Louise Coetzee is thanked for redrawing the illustrations. The following are thanked for comments and input on the keys: Seth Irish, Nil Rahola, Vincent Robert, Michael Faulde, Lizette Koekemoer and Basil Brooke. Two anonymous reviewers are thanked for comments.

Authors' contributions

The author read and approved the final manuscript.

Funding

The South African National Research Foundation (Grant No. 113300) is thanked for funding.

Availability of data and materials

Not applicable.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The author declares no competing interests.

Author details

¹ Wits Research Institute for Malaria, School of Pathology, Faculty of Health Sciences, University of the Witwatersrand, Johannesburg, South Africa. ² Centre for Emerging Zoonotic & Parasitic Diseases, National Institute for Communicable Diseases, Johannesburg, South Africa.

Received: 23 August 2019 Accepted: 29 January 2020 Published online: 13 February 2020

References

- De Meillon M. Illustrated keys to the full-grown larvae and adults of South African anopheline mosquitoes. Publ Sth Afr Inst Med Res. 1931;28:275–375.
- 2. Gillies MT, Coetzee M. A supplement to the Anophelinae of Africa South of the Sahara. Publ Sth Afr Inst Med Res. 1987;55:1–143.
- Gillies MT, De Meillon B. The Anophelinae of Africa South of the Sahara. Publ Sth Afr Inst Med Res. 1968;54:1–343.

Coetzee *Malar J* (2020) 19:70 Page 20 of 20

- Sinka ME, Bangs MJ, Manguin S, Coetzee M, Mbogo CM, Hemingway J, et al. The dominant *Anopheles* vectors of human malaria in Africa, Europe and the Middle East: occurrence data, distribution maps and bionomic précis. Parasit Vectors. 2010;3:117.
- Kyalo D, Amratia P, Mundia CW, Mbogo CM, Coetzee M, Snow RW. A geocoded inventory of anophelines in the Afrotropical Region south of the Sahara: 1898–2016. Wellcome Open Res. 2017;2:57.
- Brunhes J, Le Goff G, Geoffroy B. Afro-tropical anopheline mosquitoes Ill. Description of three new species: *Anopheles carnevalei* sp. nov., *An. hervyi* sp. nov., and *An. dualaensis* sp. nov., and resurrection of *An. rageaui* Mattingly and Adam. J Am Mosq Control Assoc. 1999;15:552–8.
- Brunhes J, Le Goff G, Boussès P. Anopheles Afrotropicaux. V. Description du male et des stades pre-imaginaux d'An. deemingi Service, 1970 et description d'An eouzani n. sp. (Diptera: Culicidae). Ann Soc Entomol Fr. 2003:39:179–85.
- Brunhes J, Le Goff G, Geoffroy B. Anopheles Afro-tropicaux. I. Descriptions d'especes nouvelles et changements de statuts taxonomiques (Diptera: Culicidae). Ann Soc Entomol Fr. 1997;33:173–83.
- Brunhes J, Le Goff G, Manga L, Geoffroy B. Anopheles Afrotropicaux. IV. Mise au point sur le complexe Anopheles moucheti, rehabilitation d'An. multicinctus et d'An. garnhami basilewskyi (Diptera: Culicidae). Ann Soc Entomol Fr. 1998;34:397–405.
- Abdulla-Khan R, Coetzee M, Hunt RH. Description of *Anopheles (Cellia)* seretsei sp. nov. from Kasane Botswana. J Am Mosq Control Assoc. 1998;14:248–52.
- Coetzee M, Segerman J, Hunt RH. Description of a new species Anopheles (Cellia) kosiensis (Diptera: Culicidae) from Zululand, South Africa. Syst Entomol. 1987;12:23–8.
- Rahola N, Makanga B, Yangari P, Jiolle D, Fontenille D, Renaud F, et al. Description of *Anopheles gabonensis*, a new species potentially involved in rodent malaria transmission in Gabon, Central Africa. Infect Genet Evol. 2014;28:628–34.
- Awono-Ambene HP, Kengne P, Simard F, Antonio-Nkondjio C, Fontenille D. Description and bionomics of *Anopheles (Cellia) ovengensis* (Diptera: Culicidae), a new malaria vector species of the *Anopheles nili* group from South Cameroon. J Med Entomol. 2004;41:561–8.
- Faulde MK, Rueda LM, Khaireh BA. First record of the Asian malaria vector *Anopheles stephensi* and its possible role in the resurgence of malaria in Djibouti, Horn of Africa. Acta Trop. 2014;139:39–43.
- Seyfarth M, Khaireh BA, Abdi AA, Bouh SM, Faulde MK. Five years following first detection of *Anopheles stephensi* (Diptera: Culicidae) in Djibouti, Horn of Africa: populations established—malaria emerging. Parasitol Res. 2019;118:725–32.
- Carter TE, Yared S, Gebresilassie A, Bonnell V, Damodaran L, Lopez K, et al. First detection of *Anopheles stephensi* Liston, 1901 (Diptera: Culicidae) in Ethiopia using molecular and morphological approaches. Acta Trop. 2018;188:180–6.

- 17. Lambert DM, Coetzee M. A dual genetical and taxonomic approach to the resolution of the mosquito taxon *Anopheles (Cellia) marshallii* (Diptera: Culicidae). Syst Entomol. 1982;7:321–32.
- Scott JA, Brogdon WG, Collins FH. Identification of single specimens of the Anopheles gambiae complex by the polymerase chain reaction. Am J Trop Med Hyg. 1993;49:520–9.
- Koekemoer LL, Kamau L, Hunt RH, Coetzee M. A cocktail polymerase chain reaction (PCR) assay to identify members of the *Anopheles funestus* (Diptera: Culicidae) group. Am J Trop Med Hyg. 2002;66:804–11.
- Coetzee M, Hunt RH, Wilkerson R, Della Torre A, Coulibaly MB, Besansky NJ. Anopheles coluzzii and Anopheles amharicus, new members of the Anopheles gambiae complex. Zootaxa. 2013;3619:246–74.
- Barrón MG, Paupy C, Rahola N, Akone-Ella O, Ngangue MF, Wilson-Bahun TA, Pombi M, Kengne P, Costantini C, Simard F, González J, Ayala D. A new species in the major malaria vector complex sheds light on reticulated species evolution. Sci Rep. 2019;9:14753.
- 22. Harbach RE, Knight KL. Taxonomists' glossary of Mosquito Anatomy. New Jersey: Plexus Publ; 1980.
- 23. Harbach RE, Knight KL. Corrections and additions to *Taxonomists' Glossary of Mosquito Anatomy*. Mosq Syst. 1981;13:201–17.
- Cumming JM, Wood DM. Adult morphology and terminology. 3. In: Kirk-Spriggs AH, Sinclair BJ, Eds Manual of Afrotropical Diptera. Vol. 1. Introductory chapters and keys to Diptera families. Suricata 4. South African National Biodiversity Institute, Pretoria; 2017. p. 89–133.
- 25. Evans AM. Mosquitoes of the Ethiopian Region. II. Anophelini Adults and Early Stages. Brit Mus (Nat Hist), London; 1938.
- Spillings BL, Brooke BD, Koekemoer LL, Chiphwanya J, Coetzee M, Hunt RH. A new species concealed by *Anopheles funestus* Giles, a major malaria vector in Africa. Am J Trop Med Hyg. 2009;81:510–5.
- Cohuet A, Simard F, Toto JC, Kengne P, Coetzee M, Fontenille D. Species identification within the *Anopheles funestus* group of malaria vectors in Cameroon and evidence for a new species. Am J Trop Med Hyg. 2003;69:200–5.
- 28. Harbach RE, Kitching IJ. The phylogeny of Anophelinae revisited: inferences about the origin and classification of *Anopheles* (Diptera: Culicidae). Zoo Scripta. 2016;45:34–47.
- 29. Snodgrass RE. The anatomical life of the mosquito. Smithsonian Institution. 1959;139:1–87.
- McAlpine JF. Morphology and terminology—adults. 2. In: McAlpine JF, Peterson BV, Shewell GE, Teskey HJ, Vockeroth JR, Wood DM, Eds. Manual of Nearctic Diptera. Vol. 1. Monograph, Research Branch, Agriculture, Canada; 1981;27:9-63.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- $\bullet\,$ thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

